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## Regulatory DNA sequences of the gene for the human catalytic telomerase subunit, and their diagnostic and therapeutic use

#### Structure and function of the chromosome ends

The genetic material of eukaryotic cells is distributed on linear chromosomes. The ends of hereditary units are termed telomeres, derived from the Greek words *telos* (end) and *meros* (part, segment). Most telomeres consist of repeats of short sequences which are mainly composed of thymine and guanine (Zakian, 1995). In all the vertebrates which have so far been investigated, the telomeres consist of the sequence TTAGGG (Meyne *et al.*, 1989).

The telomeres have a variety of important functions. They prevent the fusion of chromosomes (McClintock, 1941) and thus the formation of dicentric hereditary units. Such chromosomes having two centromeres can lead to the development of cancer due to loss of heterozygosis or duplication, or loss of genes.

In addition, telomeres serve the purpose of distinguishing intact hereditary units from damaged hereditary units. Thus, yeast cells ceased their cell division when they contained a chromosome without a telomere (Sandell and Zakian, 1993).

Telomeres fulfil another important task in association with the replication of eukaryotic cell DNA. In contrast to the circular genomes of prokaryotes, the linear chromosomes of eukaryotes cannot be completely replicated by the DNA polymerase complex. RNA primers are required to initiate DNA replication. After elimination of the RNA primers, extension of the Okazaki fragments and subsequent ligation, the newly synthesized DNA strand lacks the 5' end since the RNA primer cannot be replaced by DNA at that point. Without special protective mechanisms, the chromosomes would therefore shrink with each cell division ("end-replication problem"; Harley *et al.*, 1990). The non-coding telomere sequences presumably constitute a buffer zone for preventing the loss of genes (Sandell and Zakian, 1993).

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In addition to this, telomeres also play an import role in regulating cell ageing (Olovnikov, 1973). Human somatic cells exhibit a limited capacity for replication in culture; after a certain period of time, they become senescent. In this state, the cells no longer divide even after having been stimulated with growth factors; however, they do not die and remain metabolically active (Goldstein, 1990). Various observations support the hypothesis that a cell determines how many more times it can divide on the basis of the length of its telomeres (Allsopp *et al.*, 1992).

In summary, the telomeres consequently possess key functions in the ageing of cells, and in stabilizing the genetic material and preventing cancer.

#### The enzyme telomerase synthesizes the telomeres

As described above, organisms which possess linear chromosomes can only replicate their genome incompletely in the absence of a special protective mechanism. Most eukaryotes use a special enzyme, i.e. telomerase, for regenerating the telomere sequences. Telomerase is expressed constitutively in the single-cell organisms which have so far been investigated. On the other hand, telomerase activity has only been measured in humans in germ cells and tumour cells, whereas neighbouring somatic tissue did not contain any telomerase (Kim *et al.*, 1994).

Telomerase can also be designated functionally as terminal telomere transferase, which is located in the cell nucleus as a multiprotein complex. While the RNA moiety of human telomerase has been known for a relatively long period of time (Feng *et al.*, 1995), the catalytic subunit of this enzyme group was recently identified in a variety of organisms (Lingner *et al.*, 1997; cf. our application PCT EP/98/03468 which is likewise pending). These catalytic subunits of telomerase are strikingly homologous both among themselves and in relation to all previously known reverse transcriptases.

WO 98/14592 also describes nucleic acid and amino acid sequences of the catalytic telomerase subunit.

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#### Activation of telomerase in human tumours

It was originally only possible to demonstrate telomerase activity in humans in germ line cells and not in normal somatic cells (Hastie *et al.*, 1990; Kim *et al.*, 1994). Following the development of a more sensitive detection method (Kim *et al.*, 1994), a low telomerase activity was also detected in hematopoietic cells (Broccoli *et al.*, 1995; Counter *et al.*, 1995; Hiyama *et al.*, 1995). It is true, however, that these cells nevertheless exhibited a reduction in the telomeres (Vaziri *et al.*, 1994; Counter *et al.*, 1995). It has still not been resolved whether the quantity of enzyme in these cells is not sufficient for compensating the telomere loss or whether the telomerase activity which is measured stems from a subpopulation, e.g. incompletely differentiated CD34<sup>+</sup>38<sup>+</sup> precursor cells (Hiyama *et al.*, 1995). In order to resolve this, it would be necessary to detect telomerase activity in a single cell.

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Interestingly, however, significant telomerase activity was detected in a large number of the tumour tissues which had thus far been tested (1734/2031, 85%; Shay, 1997), whereas no activity was found in normal somatic tissue (1/196, <1%, Shay, 1997). In addition various investigations have shown that the telomeres still shrank in senescent cells which were transformed with viral oncoproteins and it was only possible to detect telomerase in the subpopulation which survived the growth crisis (Counter *et al.*, 1992). The telomeres were also stable in these immortalized cells. (Counter *et al.*, 1992). Similar findings from investigations in mice (Blasco *et al.*, 1996) support the assumption that reactivation of the telomerase is a late event in tumorigenesis.

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Based on these results, a "telomerase hypothesis" was developed which links the loss of telomere sequences and cell ageing with telomerase activity and the development of cancer. In long-lived species such as humans, the shrinking of the telomeres can be regarded as being a mechanism for suppressing tumours. Differentiated cells which do not contain any telomerase cease their cell division at a particular telomere length. If such a cell mutates, it can only form a tumour if the cell can extend its telomeres.

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Otherwise, the cell would continue to lose telomere sequences until its chromosomes became unstable and it was finally destroyed. Telomerase reactivation is presumably the main mechanism used by tumour cells to stabilize their telomeres.

It follows from these observations and considerations that it should be possible to treat tumours by inhibiting the telomerase. Conventional cancer therapies using cytostatic agents or short-wave radiation damage all the dividing cells in the body in addition to the tumour cells. However, since only germ line cells, apart from tumour cells, contain significant telomerase activity, telomerase inhibitors would attack the tumour cells more specifically and consequently elicit fewer undesirable side effects. Telomerase activity has been detected in all the tumour tissues which have so far been tested, which means that these therapeutic agents could be employed against all types of cancer. The effect of telomerase inhibitors would then set in when the telomeres of the cells had shortened to such an extent that the genome became unstable. Since tumour cells usually possess telomeres which are shorter than those of normal somatic cells, cancer cells would be the first to be eliminated by the telomerase inhibitors. By contrast, cells possessing long telomeres, such as the germ cells, would only be damaged at a much later date. Telomerase inhibitors consequently represent a potential way forward in the treatment of cancer.

It becomes possible to obtain unambiguous answers to the question of the nature and points of attack of physiological telomerase inhibitors once the manner in which expression of the telomerase gene is regulated has also been identified.

#### 25 Regulation of gene expression in eukaryotes

There are a large number of points in eukaryotic gene expression, i.e. the cellular flow of information from the DNA to the protein by way of the RNA, at which regulatory mechanisms can exert an effect. Examples of individual control steps are gene amplification, the recombination of gene loci, chromatin structure, DNA methylation, transcription, post-transcriptional modifications of mRNA, mRNA transport, translation and post-translational modifications of proteins. Studies which

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have been carried out to date indicate that control at the level of transcription initiation is of the greatest importance (Latchman, 1991).

A region which is responsible for regulating transcription, and which is designated the promoter region, is located directly upstream of the transcription start of a gene which is transcribed by RNA polymerase II. Comparison of the nucleotide sequences of promoter regions from a large number of known genes shows that particular sequence motifs occur regularly in this region. These elements include, inter alia, the TATA box, the CCAAT box and the GC box, which elements are recognized by specific proteins. The TATA box, which is located about 30 nucleotides upstream of the transcription start, is, for example, recognized by the TFIID subunit TBP ("TATA box-binding protein"), whereas particular GC-rich sequence segments are specifically bound by the transcription factor Sp1 ("specificity protein1").

The promoter can be functionally subdivided into a regulatory segment and a constitutive segment (Latchman, 1991). The constitutive control region comprises the so-called core promoter which enables transcription to be initiated correctly. This promoter contains the sequence elements which are described as UPE's (upstream promoter elements) which are necessary for efficient transcription. The regulatory control segments, which can be interlaced with the UPE's, possess sequence elements which can be involved in the signal-dependent regulation of transcription by hormones, growth factors, etc. They impart tissue-specific or cell-specific promoter properties.

DNA segments which are able to exert an influence on gene expression over relatively large distances are a characteristic feature of eukaryotic genes. These elements can be located upstream or downstream of a transcription unit, or within the unit, and can perform their function independently of their orientation. These sequence segments may reinforce (enhancers) or attenuate (silencers) promoter activity. In a similar way to the promoter regions, enhancers and silencers also accommodate several binding sites for transcription factors.

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The invention relates to the DNA sequences from the 5'-flanking region of the gene for the catalytically active human telomerase subunit and intron sequences for this gene.

The invention particularly relates to the 5'-flanking regulatory DNA sequence which contains the promoter DNA sequence for the gene for the human catalytic telomerase subunit, as depicted in Fig. 10 (SEQ ID NO 3).

The invention furthermore relates to part regions of the 5'-flanking regulatory DNA sequence, as depicted in Fig. 4 (SEQ ID NO 1), which has a regulatory effect.

Intron sequences for the gene for the human catalytic telomerase subunit, in particular those sequences which have a regulatory effect, are also part of the subject-matter of the present invention. The intron sequences according to the invention are described in detail in the context of Example 5 (cf. SEQ ID NO 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 and 20).

The invention furthermore relates to a recombinant construct which comprises the DNA sequences according to the invention, in particular the 5'-flanking DNA sequence of the gene for the human catalytic telomerase subunit, or part regions thereof.

Preference is given to recombinant constructs which, in addition to the DNA sequences according to the invention, in particular the 5'-flanking DNA sequence of the gene for the human catalytic telomerase subunit, or part regions thereof, also contain one or more additional DNA sequences which encode polypeptides or proteins.

According to a particularly preferred embodiment, these additional DNA sequences encode antineoplastic proteins.

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Particular preference is given to those antineoplastic proteins which inhibit angiogenesis directly or indirectly. Examples of these proteins are:

Plasminogen activator inhibitor (PAI-1), PAI-2, PAI-3, angiostatin, endostatin, platelet factor 4, TIMP-1, TIMP-2, TIMP-3 and leukaemia inhibitory factor (LIF).

Antineoplastic proteins which have a direct or indirect cytostatic effect on tumours are likewise particularly preferred. These proteins include, in particular:

perforin, granzyme, IL-2, IL-4, IL-12, interferons, such as IFN-α, IFN-β and IFN-γ, TNF, TNF-α, TNF-β, oncostatin M; tumour suppressor genes, such as p53, retinoblastoma.

Particular preference is furthermore given to antineoplastic proteins which, where appropriate in addition to their antineoplastic effect, stimulate inflammations and thereby contribute to the elimination of tumour cells. Examples of these proteins are:

RANTES, monocyte chemotactic and activating factor (MCAF), IL-8, macrophage inflammatory protein (MIP-1α,-β), neutrophil activating protein-2 (NAP-2), IL-3, IL-5, human leukaemia inhibitory factor (LIF), IL-7, IL-11, IL-13, GM-CSF, G-CSF and M-CSF.

Particular preference is furthermore given to antineoplastic proteins which, due to their action as enzymes, are able to convert precursors of an antineoplastic active compound into an antineoplastic active compound. Examples of these enzymes are:

herpes simplex virus thymidine kinase, varicella zoster virus thymidine kinase, bacterial nitroreductase, bacterial β-glucuronidase, plant β-glucuronidase from *Secale cereale*, human glucuronidase, human carboxypeptidase, bacterial carboxypeptidase, bacterial β-lactamase, bacterial cytosine deaminidase, human catalase and/or phosphatase, human alkaline phosphatase, type 5 acid phosphatase, human

lysooxidase, human acid D-aminooxidase, human glutathione peroxidase, human eosinophil peroxidase and human thyroid peroxidase.

The abovementioned recombinant constructs can also contain DNA sequences which encode factor VIII or factor IX, or part fragments thereof. These DNA sequences also include other blood clotting factors.

The abovementioned recombinant constructs can also contain DNA sequences which encode a reporter protein. Examples of these reporter proteins are:

Chloramphenicol acetyl transferase (CAT), glow-worm luciferase (LUC), \(\beta\)-galactosidase (\(\beta\)-Gal), secreted alkaline phosphatase (SEAP), human growth hormone (hGH), \(\beta\)-glucuronidase (GUS), green-fluorescing protein (GFP), and all the variants

derived therefrom, aquarin and obelin.

Recombinant constructs according to the invention can also contain DNA which encodes the human catalytic telomerase subunit and its variants and fragments in the antisense orientation. Where appropriate, these constructs can also contain other protein subunits of the human telomerase and the telomerase RNA component in the antisense orientation.

The recombinant constructs can, in addition to the DNA which encodes the human catalytic telomerase subunit, and its variants and fragments, also contain other protein subunits of the human telomerase and the telomerase RNA component.

The invention furthermore relates to a vector which contains the abovementioned DNA sequences according to the invention, in particular the 5'-flanking DNA sequences and also one or more of the other DNA sequences mentioned above.

The preferred vector for these constructs is a virus, for example a retrovirus, an adenovirus, an adeno-associated virus, a herpes simplex virus, a vaccina virus, a lentiviral virus, a Sindbis virus and a Semliki forest virus.

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Preference is also given to using plasmids as vectors.

The invention furthermore relates to pharmaceutical preparations which comprise recombinant constructs or vectors according to the invention; for example a preparation in a colloidal dispersion system.

Examples of suitable colloidal dispersion systems are liposomes or polylysine ligands.

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The preparations of the constructs or vectors according to the invention in colloidal dispersion systems can be supplemented with a ligand which binds to the membrane structures of tumour cells. Such a ligand can, for example, be attached to the construct or the vector or else be a component of the liposome structure.

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Suitable ligands are, in particular, polyclonal or monoclonal antibodies, or antibody fragments thereof, which bind, by their variable domains, to the membrane structures of tumour cells, or substances carrying mannose terminally, cytokines or growth factors, or fragments or part sequences thereof, which bind to receptors on tumour cells.

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Examples of corresponding membrane structures are receptors for a cytokine or a growth factor, such as IL-1, EGF, PDGF, VEGF, TGF β, insulin or insulin-like growth factor (ILGF), or adhesion molecules, such as SLeX, LFA-1, MAC-1, LECAM-1 or VLA-4, or the mannose-6-phosphate receptor.

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The present invention includes pharmaceutical preparations which, in addition to the vector constructs according to the invention, can also comprise non-toxic, inert, pharmaceutically suitable excipients. It is possible to conceive of administering (e.g. intravenously, intraarterially, intramuscularly, subcutaneously, intradermally, anally, vaginally, nasally, transdermally, intraperitoneally, as an aerosol or orally) these preparations at the site of a tumour or administering them systemically.

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The vector constructs according to the invention can be employed in gene therapy.

The invention furthermore relates to a recombinant host cell, in particular a recombinant eukaryotic host cell, which harbours the above-described constructs or vectors.

The invention furthermore relates to a process for identifying substances which affect the promoter activity, silencer activity or enhancer activity of the catalytic telomerase subunit, with this process comprising the following steps:

- A. adding a candidate substance to a host cell which harbours the regulatory DNA sequence according to the invention, in particular the 5'-flanking regulatory DNA sequence for the gene for the human catalytic telomerase subunit, or a part region thereof which has a regulatory effect, which sequence or part region is functionally linked to a reporter gene, and
- B. measuring the effect of the substance on expression of the reporter gene.
- The process can be employed for identifying substances which increase the promoter activity, silencer activity or enhancer activity of the catalytic telomerase subunit.

The process can furthermore be employed for identifying substances which inhibit the promoter activity, silencer activity or enhancer activator of the catalytic telomerase subunit.

The invention furthermore relates to a process for identifying factors which bind specifically to fragments of the DNA fragments according to the invention, in particular the 5'-flanking regulatory DNA sequence of the catalytic telomerase subunit. This method comprises screening an expression cDNA library using the above-described DNA sequence, or subfragments of widely differing length, as the probe.

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The above-described constructs or vectors can also be used for preparing transgenic animals.

- The invention furthermore relates to a process for detecting telomerase-associated conditions in a patient, which process comprises the following steps:
  - A. incubating a construct or vector, which contains the DNA sequence according to the invention, in particular the 5'-flanking regulatory DNA sequence for the gene for the human catalytic telomerase subunit, or a part region thereof having a regulatory effect, and a reporter gene, with body fluids or cell samples,
  - B. detecting the activity of the reporter gene in order to obtain a diagnostic value; and
  - C. comparing the diagnosic value with standard values for the reporter gene construct in standardized normal cells or body fluids of the same type as the test sample;

The detection of diagnostic values which are higher or lower than the standard comparative values indicates a telomerase-associated condition, which in turn indicates a pathogenic condition.

- 25 Explanation of the figures:
  - Fig. 1: Southern blot analysis using genomic DNA from various species

A: Photograph of an ethidium bromide-stained 0.7% agarose gel containing approximately 4 μg of Eco RI-cut genomic DNA. Track 1 contains Hind III-cut λ DNA as size markers (23.5, 9.4, 6.7, 4.4, 2.3, 2.0 and 0.6 kb). Tracks 2 to 10 contain human, rhesus monkey. Sprague

Dawley rat, BALB/c mouse, dog, bovine, rabbit, chicken and yeast (Saccharomyces cerevisiae) genomic DNA.

B: Autoradiogram, corresponding to Fig.1 A, of a Southern blot analysis in which radioactively labelled hTC-cDNA probe of about 720 bp in length is used for the hybridization.

Fig. 2: Restriction analysis of the recombinant  $\lambda$  DNA of the phage clone P12, which hybridizes with a probe from the 5' region of the hTC cDNA.

The figure shows a photograph of an ethidium bromide-stained 0.4% agarose gel. Tracks 1 and 2 contain Eco RI/Hind III-cut  $\lambda$  DNA and a 1 kb ladder from Gibco as size markers. Tracks 3 - 7 each contain 250 ng of the DNA from the recombinant phage which has been cut with Bam HI (track 3), Eco RI (track 4), Sal I (track 5), Xho I (track 6) and Sac I (track 7). The arrows mark the two  $\lambda$  arms of the vector EMBL3 Sp6/T7.

Fig. 3: Restriction analysis and Southern blot analysis of the recombinant  $\lambda$  DNA of the phage clone which hybridizes with a probe from the 5' region of the hTC cDNA.

A: The figure shows a photograph of an ethidium bromide-stained 0.8% agarose gel. Tracks 1 and 15 contain a 1 kb ladder from Gibco as size markers. Tracks 2 to 14 each contain 250 ng of cut λ DNA from the recombinant phage clone. The following enzymes were employed: track 2: Sac I, track 3: Xho I, track 4: Xho I, Xba I, track 5: Sac I, Xho I, track 6: Sal I, Xho I, Xba I, track 7: Sac I, Xho I, Xba I, track 8: Sac I, Sal I, Xba I, track 9: Sac I, Sal I, BamH I, track 10: Sac I, Sal I, Xho I, track 11: Not I, track 12: Sma I, track 13: empty, track 14: not digested.

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B: Autoradiogram, corresponding to Fig. 3 A, of a Southern blot analysis. A 5'-hTC cDNA fragment of about 420 bp in length was used as the probe for the hybridization.

- Fig. 4: Partial DNA sequence of the 5'-flanking region and of the promoter of the gene for the human catalytic telomerase subunit. The ATG start codon in the sequence is printed in bold. The depicted sequence corresponds to SEQ ID NO 1.
- Fig. 5: Use of primer extension analysis to identify the transcription start.

The figure shows an autoradiogram of a denaturing polyacrylamide gel which was selected for depicting a primer extension analysis. An oligonucleotide having the sequence 5'GTTAAGTTGTAGCTTACACTGGTTCTC 3' was used as the primer. The primer extension reaction was loaded in track 1. Tracks G, A, T and C constitute the sequence reactions using the same primer and the corresponding dideoxynucleotides. The thick arrow marks the main transcription start while the thin arrows point to three subsidiary transcription start points.

- Fig. 6: cDNA sequence of the human catalytic telomerase subunit (hTC; cf. our pending application PCT/EP/98/03468). The depicted sequence corresponds to SEQ ID NO 2.
- Fig. 7: Structural organization and restriction map of the human hTC gene and its 5'-flanking and 3'-flanking regions.

Exons are shown as consecutively numbered rectangles which are filledin in black, and introns are shown as regions which are not filled in. Untranslated sequence segments in the exons are hatched. Translation starts in exon 1 and ends in exon 16. Restriction enzyme cleavage sites

are marked as follows: S, SacI; X, XhoI. The relative arrangement of the five phage clones (P2, P3, P5, P12, P17), and of the product from the genome walking, are shown by thin lines. As the dots indicate, the sequence of intron 16 has only been partly deciphered.

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Fig. 8: HTL splice variants.

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A: Diagrammatic structure of the hTC mRNA splice variants. The complete hTC mRNA is depicted as a rectangle with a grey background in the upper region of the figure. The 16 exons are depicted in accordance with their size. The translation start (ATG) and the stop codon, and also the telomerase-specific T motif, and the seven RT motifs, are all shown. The hTC variants are subdivided into deletion and insertion variants. The missing exon sequences are marked in the deletions. The insertions are shown by additional white rectangles. The sizes and origins of the inserted sequences are given. Newly formed stop codons are marked. The size of the insertion in variant INS2 is unknown.

B: Exon-intron transitions in the hTC splice variants. Unspliced 5'flanking and 3'-flanking sequences are shown as white rectangles. The origins of the exon and intron sequences are given. Intron and exon sequences are shown in small letters and large letters, respectively. The donor and acceptor sequences in the splice sites are underlaid as grey rectangles, and their exon and intron origins are also given.

Fig. 9: Identification of the transcription start by means of RT-PCR analysis.

> The RT-PCR was carried out using a cDNA library prepared from HL 60 cells and genomic DNA as the positive control. A common 3' primer hybridizes to a region of the exon 1 sequence. The positions of the different 5' primers in the coding region or the 5'-flanking region are given. In the negative control, no template DNA was added to the PCR reaction. M: DNA size marker.

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Fig. 10: Nucleotide sequence and structural features of the hTC promoter.

The figure depicts 11273 bp of the 5'-flanking hTC gene sequence, beginning with the translation start codon ATG (+1). The putative region of the translation start is underlined. Possible regulatory sequence segments within the 4000 bp upstream of the translation start are ringed. The depicted sequence corresponds to SEQ ID NO 3.

Fig. 11: Activity of the hTC promoter in HEK-293 cells.

The first 5000 bp of the 5'-flanking hTC gene region are shown diagrammatically in the upper part of the figure. The ATG start codon is picked out. CpG-rich islands are marked by grey rectangles. The sizes of the hTC promoter-luciferase construct are shown on the left-hand side of the figure. The promoterless pGL2 basic construct and the SV40 promoter construct pGL2-Pro were used as controls in each transfection. The relative luciferase activities of the different promoter constructs in HEK cells are shown as continuous bars on the right-hand side of the figure. The standard deviation is indicated. The numerical values represent the average of two independent experiments which were carried out in duplicate.

Tab. 1: Exon-intron transitions in the hTC gene

The table lists the nucleotide sequences at the 3' and 5' splice transitions of the hTC gene. The consensus sequences for donor and acceptor sequences (AG and GT) are underlaid with grey rectangles. The table shows the intron sequences (small letters) and exon sequences (large letters) which flank the splice acceptor and donor sites. The sizes of the exons and introns are given in bp.

Tab. 2: Potential binding sites for DNA-binding factors in the nucleotide sequence of intron 2

The search for possible DNA-binding factors (e.g. transcription factors) was carried out using the "find pattern" algorithm from the Genetics Computer Group (Madison, USA) GCG sequence analysis program package. The table lists the abbreviations of the DNA-binding factors which were identified and their location in intron 2.

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Tab. 1

Intron	3' Acceptor Sequence			S	5' Donor Sequence			
Exon   Exon   Pp   Exon   Pp   Exon   Pp   Exon   No.								
No.	Intron	Exon	Exon	dq	Exon	Intron	Intron	ďq
GITTCAGGCAGCGCTGCGT         1         281         CGCCCCTCCTCCGCCAGG           GTGTCCTGCCTGAAGGAGC         3         196         TGGAAGCATTGGAATCAG           ACAGCACTTGAAGAGGTG         4         181         GTTCCGCAGAAAAGAGG           ACAGCACTTGAAGAGGTG         5         180         TGAGCTGTTTGTCAAG           GCGAGCGTCTCACCTCGA         5         180         TGAGCTGTACTTTGTCAAG           GTCCTACCTTGACAGACC         7         96         TGCGTGTACTTTGTCAAG           GTCTCTACCTTGACAGACC         7         96         TGCGTGCTCTACGAGGCAC           ACCTCTACGTCCAGTGCCAG         8         8         CCGTGCGCATCGAGCAG           GTCTTACGTCCAGTGCTG         10         72         ACGCGAAAACCTTCCTCAG           GTCTTCTACGTCCAGTGTG         11         189         TGCGGAACTTCCTCAG           GTTTCACGGACTCCAGTG         12         127         CCTGTTTCTGGATTTGCAG           GTTACACGCATGTGTGTG         13         6         TCTGCTGCTGCAGCGTACAG           GTTACACGCATGTGTGTG         12         127         CCTGTTTCTGGATTTGCAG           GTTACACGCATGTGTGTGTG         14         125         CTGAAAGCCTCAGGACG           GTTACACGCATGTGTGTGTG         14         125         CTGAAAGCCTCAGGACGCAAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACCAGAACA			No.				No.	- tribberthisternen
GTTTCAGGCAGCGCTGCGT         1         281         CGCCCCTCCTCCGCCAG           GTGTCCTGCCTGAGGAGC         2         1354         TGGCTGCGCAGAGCCCAG           GGGTTGGCTGTTCCGGC         3         196         TGAAAGCATTGGAATCAG           ACAGCACTTGAAGAGGGTG         4         181         GTTCCGCAGAGAAAGAGG           GCCGAGCGTCTCACCTCGA         5         180         TGAGCTGTACTTGTCAAG           GCGGATGTGACGAGCGGT         6         156         CAAGGCTTTGTCAAG           GTCCTACCTTGACATGAGG         8         8         CCGTGCCTTCAAGGGCAG           AGCTCCTCCTCCTGATTGGTG         10         72         ACGCGGAATTCGCGGGGAAA           GTCTACCTGCTGGTTTGGTG         10         72         ACGCGAAAACCTCCAG           GTCTACCTGGTCCAGGTTC         11         189         TGCAGAGCGATTCCTCAG           GTCTACCTGGTCCAGGGTTC         11         189         TGCAGAGCGATTCCTCAG           GTGAACAGCCTCCAGACGGTT         12         127         CCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGACGCACAAGAACC         12         TGCAGAGCGCTACCAGGCAAA           GTGAACAGCCTCCAGACGCAAA         13         6         TCCTGCTGCTGCAGACGACAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACCAAGAACACAAGAACAAC	)  -							
GTGTCTGCCTGAAGGAGC         2         1354         TGGCTGCGCAGGAGCCAGG           GGGTTGGCTGTTCCGGC         3         196         TGCAAAGCATTGGAATCAG           ACAGCACTTGAAGAGGTG         4         181         GTTCCGCAGAGAAACAG           GCCGAGCTCTCACCTCGA         5         180         TGAGCTGTACTTGTCAAG           GTCTCTACCTTGACAGACC         7         96         TGCGGTGTTGTCAAG           GTCTCTACCTTGACAGACC         8         8         CCGTGCGTCAGGGGCAA           AGCTCTCCTGAATGAG         9         114         CGGGGGTTCAGGGGCAA           GTCTCTACGTCGAGTGCCAG         9         114         CGGGGATTCGAGGGCAA           GTCTCTACGTCGAGTGCCAG         9         114         CGGGGATACTCCAGGGCAA           GTCTCTCCGGAGTTGGTG         10         72         ACGCGAAAACCTCCTCAGGGCAA           GTGAACAGCCTCCAGGGGTC         11         189         TGCAGAGCTCCTCAG           GTGAACAGCCTCCAGACCG         13         62         TCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGACCG         13         62         TCTGCTTCTGCAGACCAG           GTTTCACCTGCAGGGCCAA         14         125         TCTGCTTCTGCAGACCAG           GTTTCACCTGCAGGGCCAA         15         TCTGCTTTCTGGATTTGAAAAACTCCAGAGACCACAGACACACAC	5' flanking region	GTTTCAGGCAGCGCTGCGT	7	281	CGCCCCTCCTTCCGCCAG	gtgggcctccccggggtcg	н	104
GGGTTGGCTGTCCGGC         3         196         TGCAAAGCATTGGAATCAG           ACAGCACTTGAAGAGGTG         4         181         GTTCCGCAGAGAAAGAGG           GCCGAGCGTCTCACCTCGA         5         180         TGAGCTGTACTTTGTCAAG           GTGGATGTGCTTGACAGGCCAC         7         96         TGAGGCTTCAAGAGCCAC           GTCTCTACCTTGACATGAGG         8         8         CCGTGCGCTCATCGAGCAC           AGCTCCTCCCTGCATTGGTG         9         114         CGGGGATTCAGGGCAC           GTCTTACCTTGCTGTGTG         10         72         ACGCGGATTTGCTGG           GCTGCTCCTGGTCCTGTGTGT         11         189         TGCAGAGCGTCCTGG           GTGTTCAGGTCCATC         12         127         CCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGCG         13         62         TCTGAAACACCTCCAGG           GTTTCAGGCTTGGGGCCCAA         14         125         CTGAAAGCCACAGACCAG           GTTTCACGCTGGGGCCCAA         15         125         CTGAAAGCCACCAGCACAGACACAGACACACACACACACA	cagggcgcttcccccgdag	GIGICCIGCCIGAAGGAGC	7	1354	TGGCTGCGCAGGAGCCCAG	gtgaggaggtggtggccgt	8	8616
ACAGCACTTGAAGAGGGTG         4         181         GTTCCGCAGAAAAAGAGG           GCCGAGCGTCTCACCTCGA         5         180         TGAGCTGTACTTTGTCAAG           GTGCATGTGACGGCGCT         6         156         CAAGGCCTTCAAGAGG           GTCTCTACCTTGACAGGC         7         96         TGCGGCGTCATCGAGGCAA           AGCTCCTCCTCAATGAG         8         86         CCGTGCGCATCAGGGGAA           GTCCTCTCCTCAATGAG         9         114         CGGGGATTCGGGGGAAA           GTCTCTACCTCAGTGCTA         9         114         CGGGGATTCGGGGGAAA           GTCTCTACCTCAGTGTG         10         72         ACGCGAAAACCTCCTCAG           GACCTGGTCCAGGTGTC         11         189         TGCAGAGCGATCTCCAG           GACCTGGTCCAGGCTCCATC         12         127         CCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGACG         13         62         TCTGTTTCTGGATTTGCAG           GTTTCACGCTGGGGCCAA         15         125         CTGAAAAGCCTCAGGACAG           GTTTCACGCTGGGGCCAA         15         125         CTGAAACAGCCTCAGGACAG           GGATTCACGCTGGGGCCAA         15         138         CTGGGGTCACAGAAAACCTTCAGGACAG           GGATTCACGCTGAGGCCAAA         15         138         CTGGGGCCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA	catgtccttctcgtttaag	GGGTTGGCTGTGTTCCGGC	m	196	TGCAAAGCATTGGAATCAG	gtactgtatccccacgcca	m	2089
GCCGAGCGTCTCACCTCGA         5         180         TGAGCTGTACTTTGTCAAG           GTCGATGTGACGGGCGCT         6         156         CAAGGCCTTCAAGAGCAC           GTCTCTACCTTGACAGACC         7         96         TGCCGTCGTCATCGAGCAG           AGCTCCTCCCTGAATGAGG         8         86         CCGTGCGCATCGAGGCAA           GTCCTACGTCCAGTGCCAG         9         114         CGGGGATTCGGGGCAA           GCTGCTCCTGCGTTTGGTG         10         72         ACGCGAAAACCTTCCTCAG           GACCTTGGTCCGAGGTGTC         11         189         TGCAGACCTCCAG           GACCTTGGTCCGAGGTGTC         11         189         TGCAGACCTCCAG           GTGAACAGCCTCCAGC         13         62         TCTGTTTCTGATTTGCAG           GTGAACAGCCTCCAGGCCAA         14         125         CTGAAAGCCAAGAACAG           GGATGTCGCGGGGCCAA         15         138         CTGGGGCTACAGGACAG           GCAGACCTGAGGCCAA         15         138         CTGGAAACAGCCACAGGACAG           GCAGACGCAGGAGCGAA         15         138         CTGGAAACACCACAGGACAG           GCAGAACGCAGGAGCAA         16         664         TTTTTCAGGTTTGAAAAA	gagggctctctattgcag	ACAGCACTTGAAGAGGGTG	4	181	GTTCCGCAGAGAAAGAGG	gtggctgtgctttggttta	4	687
GTGGATGTGACGGCGCGT         6         156         CAAGGCCTTCAAGAGCCAC           GTCTCTACCTTGACAGACC         7         96         TGCCGTCGTCATCGAGGCAA           AGCTCCTCCCTGAATGAGG         8         86         CCGTGCGCATCAGGGGCAA           GTCCTACGTCCAGTGCCAG         9         114         CGGGGATTCGGCGAGGGCAA           GTCTGCTCCTGCGTTGGTG         10         72         ACGCGAAAACCTTCCTCAG           GACCTGGTCCAGGTGTC         11         189         TGCAGAGCGACTCCAGG           CTATGCCCGGACCTCCATC         12         127         CCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGCG         13         62         TCTGCTGCAGGCGACAGG           GTTTCACGCATGTGCTG         14         125         CTGAAAGCCAACGAACGCAGG           GGATGTCGCTGGGGCCAA         15         138         CTGGGGTCACTCAGGACAG           GGATGTCGCTGGGGCCAA         15         138         CTGGGGTCACTCAGGACAG           GCCAGACGCTGAGTCG         16         664         TTTTTCAGTTTTGAAAAA	cccatgctgtccccgccag	GCCGAGCGTCTCACCTCGA	ប	180	TGAGCTGTACTTTGTCAAG	gtgggtgccggggacccc	S	494
GTCTCTACCTTGACAGACC 7 96 TGCCGTCGTCATCGAGCAG AGCTCCTCCCTGAATGAGG 8 86 CCGTGCGCATCAGGGGCAA GTCCTACGTCCTGCGTTTGGTG 10 72 ACGCGAAACCTTCCTCAG GACCCTGGTCCAGTGTC 11 189 TGCAGAGCGACTACTCCAG GACCCTGGTCCAGTGTC 12 127 CCTGTTTCTGGATTTGCAG GTGAACAGCCTCCAGAGGGTTC 13 62 TCCTGCTGCAGGCGTACAG GTTTCACGCATGTGTG 14 125 CTGGGGTCACTCAGG GGATGTCGCTGGGGCCAA 15 138 CTGGGGTCACTCAGGACAG CCCAGACGCTGAGTCG 16 664 TTTTTCAGTTTTGAAAAA	ctcgcctccactcacadag	GTGGATGTGACGGGCGCGT	9	156	CAAGGCCTTCAAGAGCCAC	gtaaggttcacgtgtgata	9	>4660
AGCTCCTCCTGAATGAGG 8 86 CCGTGCGCATCAGGGGCAA GTCCTACGTCGCCCAGG GCTGCTCCTGCTTGGTG 10 72 ACGCGAAAACCTTCCTCAG GACCCTGGTCCGAGGTGTC 11 189 TGCAGAGCGACTACTCCAG CTATGCCCGGACTCCATC 12 127 CCTGTTCTGGATTTGCAG GTGAACAGCCTCCAGACG 13 62 TCCTGCTGCAGGCGTACAG GTTTCACGCATGTGCTG 14 125 CTGAAAGCCAAGAACGCAG GGATGTCGCTGGGGCCAA 15 138 CTGGGGTCACTCAGGACAG CCCAGACGCCGCAAAAAAAA  CCCAGACGCCGCAGACGG 16 664 TTTTCAGTTTTGAAAAA	ccetctcctgccggcag	GICICIACCIIGACAGACC	7	96	TGCCGTCGTCATCGAGCAG	gtctgggcactgcctgca	7	980
GTCCTACGTCCAGTGCCAG  GTCCTACGTCCAGTGCCAG  GCTGCTCCTGCGTTTGGTG  10 72 ACGCGAAAACCTTCCTCAG  GACCTGGTCCAGGTGTC  11 189 TGCAGAGCGACTACTCCAG  CTATGCCCGGACCTCCATC  12 127 CCTGTTTCTGGATTTGCAG  GTGAACAGCCTCCAGACG  13 62 TCCTGCTGCAGCAGACGCAG  GTTTCACGCATGTGTGCTG  14 125 CTGAAAGCCAAGAACGCAG  GGATGTCGCTGGGGCCAA  15 138 CTGGGGTCACTCAGGACAG  CCCAGACGCAGGTCG  16 664 TTTTCAGTTTTGAAAAA	ctcccgtctgctttcgcag	AGCTCCTCCTGAATGAGG	80	98	CCGTGCGCATCAGGGGCAA	gtgagtcaggtggccaggt	80	2485
GCTGCTCCTGCGTTTGGTG         10         72         ACGCGAAAACCTTCCTCAG           GACCCTGGTCCGAGGTGTC         11         189         TGCAGAGCGACTACTCCAG           CTATGCCCGGACCTCCATC         12         127         CCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGACGG         13         62         TCTGGTGCAGCTACAG           GTTTCACGCATGTGCTG         14         125         CTGAAAGCCAAGAACGCAG           GGATGTCGCTGGGGCCAA         15         138         CTGGGGTCACTCAGGACAG           CCCAGACGCAGGTCG         16         664         TTTTTCAGTTTTGAAAAAA	ctgtgtcttcccgcccdag	GICCIACGICCAGIGCCAG	o	114	CGGGGATTCGGCGGGACGG	gtgaggcctcctcttcccc	on	1984
GACCCTGGTCCGAGGTGTC         11         189         TGCAGAGCGACTACTCCAG           CTATGCCCGGACCTCCATC         12         127         CCTGTTTCTGGATTTGCAG           GTGAACAGCCTCCAGACG         13         62         TCCTGCTGCAGGCTACAG           GTTTCACGCATGTGCTG         14         125         CTGAAAGCCAAGAACGCAG           GGATGTCGCTGGGGCCAA         15         138         CTGGGGTCACTCAGGACAG           CCCAGACGCAGGTCGAGTCG         16         664         TTTTTCAGTTTTGAAAAAA	gtattttcccttattttag	GCTGCTCCTGCGTTTGGTG	10	72	ACGCGAAAACCTTCCTCAG	gtgaggcccgtgccgtgtg	10	1871
CTATGCCCGGACCTCCATC 12 127 CCTGTTTCTGGATTTGCAG GTGAACAGCCTCCAGACGG 13 62 TCCTGCTGCAGCGTACAG GTTTCACGCATGTGTGCTG 14 125 CTGAAAGCCAAGAACGCAG GGATGTCGCTGGGGCCAA 15 138 CTGGGGTCACTCAGGACAG CCCAGACGCAGCTGAGTCG 16 664 TTTTCAGTTTTGAAAAA	cattgcccctctgccttag	GACCCIGGICCGAGGIGIC	11	189	TGCAGAGCGACTACTCCAG	gtgagcgcacctggccgga	11	3801
GTGAACAGCCTCCAGACGG 13 62 TCCTGCTGCAGGCGTACAG GTTTCACGCATGTGCTG 14 125 CTGAAAGCCAAGAACGCAG GGATGTCGCTGGGGCCAA 15 138 CTGGGGTCACTCAGGACAG CCCAGACGCAGCTGAGTCG 16 664 TTTTCAGTTTTGAAAAA	attccccctgtgtctdag	CTATGCCCGGACCTCCATC	12	127	CCTGTTTCTGGATTTGCAG	gtgagcaggctgatggtca	12	880
GITTCACGCATGTGCTG 14 125 CTGAAAGCCAAGAACGCAG GGATGTCGCTGGGGCCAA 15 138 CTGGGGTCACTCAGACAG CCCAGACGCAGCTGAGTCG 16 664 ITTTCAGTTTTGAAAAA	tctttcttggcgactctag	GTGAACAGCCTCCAGACGG	13	62	rccrecrecaegceracae	gtgagccgccaccaagggg	13	3187
GGATGTCGCTGGGGCCAA 15 138 CTGGGGTCACTCAGGACAG CCCAGACGCAGCTGAGTCG 16 664 ITTTCAGTTTTGAAAAA	ctgtccgccatcctctag	GTTTCACGCATGTGTGCTG	14	125	CTGAAAGCCAAGAACGCAG	gtatgtgcaggtgcctggc	14	781
CCCAGACGCAGCTGAGTCG 16 664 ITTTCAGTTTTGAAAAA 3.	agcetetgttttccccdag	GGATGTCGCTGGGGGCCAA	15	138	CTGGGGTCACTCAGGACAG	gcaagtgtgggtggaggcc	15	536
	totgattttggccccgdag	CCCAGACGCAGCTGAGTCG	16	664	TITITCAGITITGAAAAA	3' flanking region		

Tab. 2

Factors	Location in intron 2
C/EBP	2925
CRE.2	2749
Spl	2378, 4094, 4526, 4787, 4835, 4995
AP-2 CS3	5099
AP-2 CS4	2213, 3699, 4667, 5878, 5938, 6059, 6180, 6496
AP-2 CS5	5350, 5798, 5880, 5940, 6061, 6182, 6375, 6498
PEA3	934, 2505
P53	2125
GR uteroglobin	848, 1487, 2956
PR uteroglobin	3331 -
Zeste-white	1577, 1619, 1703, 1745, 1787, 1829, 1871, 1913, 1955,
	1997, 2039, 2081, 3518, 3709, 4765, 5014, 5055
GRE	846
MyoD-MCK right	447, 509, 558, 1370, 1595, 1900, 2028, 2099, 4557
site/rev	
MyoD-MCK left site	108, 118, 453, 1566, 1608, 1692, 1734, 1818, 1902,
	1986, 2372, 2460, 2720, 3491, 5030
Ets-1 CS	6408
API	3784, 4406
CREB	2801
GATA-1	839, 1390, 3154
с-Мус	108, 118, 453, 1566, 1608, 1692, 1734, 1818, 1902,
	1986, 2372, 2460, 2720, 3491, 5030
CACCC site	991
CCAAT site	1224
CCAC box	992
CAAT site	463, 2395
Rb site	992, 4663
TATA	3650
CDEI	106, 1564, 1606, 1690, 1732, 1816, 1900, 1984

#### **Examples**

The human gene for the catalytic telomerase subunit (ghTC), and the regions of this gene located 5' and 3', were cloned, while the start point for transcription was determined, potential binding sites for DNA-binding proteins were identified and active promoter fragments were highlighted. The sequence of the hTC cDNA (Fig. 6) has already been reported in our application PCT/EP/98/03468, which is also pending. Unless otherwise mentioned, all the data refer to the position of the cDNA in this sequence.

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#### Example 1

A genomic Southern blot analysis was used to determine whether ghTC constitutes a single gene in the human genome or whether there exist several loci for the hTC gene and possibly also ghTC pseudogenes.

In order to do this, a commercially available zoo blot from Clontech was subjected to Southern blot analysis. This blot contains 4 µg of Eco RI-cut genomic DNA from nine different species (human, monkey, rat, mouse, dog, bovine, rabbit, chicken and yeast). With the exception of yeast, chicken and human, the DNA was isolated from kidney tissue. The human genomic DNA was isolated from placenta and the chicken genomic DNA was purified from liver tissue. An hTC cDNA fragment of about 720 bp in length, which was isolated from hTC cDNA, variant Del2 (position 1685 to 2349 plus 2531 to 2590 in Fig. 6 [deletion 2; cf. Example 5 in Fig. 8]), was used as the radioactively labelled probe in the autoradiogram in Fig. 1. The experimental conditions for the blot hybridization and washing steps were taken from Ausubel *et al.* (1987).

In the case of the human DNA, the probe recognizes two specific DNA fragments. The smaller Eco RI fragment, of from about 1.5 to 1.8 kb in length, probably originates from two Eco RI cleavage sites in an intron in the ghTC DNA. On the

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basis of this result, it is to be assumed that only one single ghTC gene is present in the human genome.

#### Example 2

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In order to isolate the 5' flanking hTC gene sequence, approx. 1.5 x 10<sup>6</sup> phages from a human genomic placenta gene library (EMBL 3 SP6/T7 from Clontech, order number HL1067j) were hybridized on nitrocellulose filters (0.45 μm; from Schleicher and Schuell), in accordance with the manufacturer's instructions, with a radioactively labelled 5'-hTC cDNA fragment of about 500 bp in length (position 839 to 1345 in Fig. 6). The nitrocellulose filters were firstly incubated, at 42°C for two hours, in 2 x SSC (0.3 M NaCl; 0.5 M Tris-HCl, pH 8.0) and then in a prehybridization solution (50% formamide; 5 x SSPE, pH 7.4; 5 x Denhard's solution; 0.25% SDS; 100 μg of herring sperm DNA/ml). For the overnight hybridization, the prehybridization solution was supplemented with 1.5 x 10<sup>6</sup> cpm of denatured, radioactively labelled probe/ml of solution. Nonspecifically bound radioactive DNA was removed under stringent conditions, i.e. by means of three five-minute steps of washing with 2 x SSC; 0.1% SDS at from 55 to 65°C. The filters were evaluated by autoradiography.

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The phage clones which were identified in this primary investigation were purified (Ausubel et al. (1987)). In subsequent analyses, one phage clone, i.e. P12 turned out to be potentially positive. A  $\lambda$  DNA preparation carried out on this phage (Ausubel et al. (1987)), and the subsequent restriction digestion with enzymes which release the genomic insert in fragments, showed that this phage clone contains an insert of approx. 15 kb in the vector (Fig. 2).

In order to isolate the complete hTC gene sequence, in each case from 1 to 1.5 x 10<sup>6</sup> phages were screened, in independent experiments, with in each case different radioactively labelled probes, as described above.

The phage clones which were identified in these primary investigations, and which were positive for the corresponding probes, were purified. The phage clone P17 was found to contain an hTC cDNA fragment of about 250 bp in length (position 1787 to 2040 in Fig. 6). The phage clone P2 was identified as containing an hTC cDNA fragment of about 740 bp in length (position 1685 to 2349 plus 2531 to 2607 in Fig. 6 [deletion 2; cf. Example 5]). The phage clones P3 and P5 were found to contain a 3' hTC cDNA fragment of 420 bp in length (position 3047 to 3470 in Fig. 6). After the  $\lambda$  DNA had been prepared from these phages, and subsequently subjected to restriction digestion with enzymes which release the genomic insert in fragments, the inserts were subcloned into plasmids (Example 4).

#### Example 3

In order to investigate whether the 5' end of the hTC cDNA was also present in the insert in the recombinant phage clone P12, the  $\lambda$  DNA from this clone was hybridized, in a Southern blot analysis, with a radiactively labelled hTC cDNA fragment of about 440 bp in length (position 1 to 440 in Fig. 6) from the extreme 5' region (Fig. 3).

Since the isolated  $\lambda$  DNA from the positive clone also hybridizes with the extreme 5' end of the hTC cDNA, this phage probably also contains the 5' sequence region flanking the ATG start codon.

#### Example 4

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In order to subclone the entire 15 kb insert in the positive phage clone P12 in the form of subfragments, and subsequently to sequence these fragments, restriction endonucleases which, on the one hand, release the entire insert from EMBL3 Sp6/T7 (cf. Example 2) and, in addition, cut within the insert, were selected for digesting the DNA.

In all, two Xho I subfragments, of about 8.3 and about 6.5 kb in length, respectively, and three Sac I subfragments, of about 8.5, about 3.5 and about 3 kb in length, respectively, were subcloned into the pBluescript KS(+) vector (from Stratagene). The 5123 bp 5'-flanking nucleotide sequence of the ghTC gene region, starting from the ATG start codon, was determined by analysing the sequences of these fragments (Fig. 4; corresponding to SEQ ID NO 1). Fig. 4 depicts the first 5123 bp (starting from the ATG start codon). Fig. 10 depicts the entire cloned 5' sequence (corresponding to SEQ ID NO 3).

In order to subclone the entire insert, of approx. 14.6 kb in size, in phage clone P17 in the form of subfragments, restriction endonucleases which, on the one hand, release the entire insert from EMLB3 Sp6/T7 and, in addition, cut a few times within the insert, were selected for digesting the DNA. Three XhoI/BamHI fragments, of 7.1 kb, 4.2 kb and 1.5 kb in size, respectively, and one BamHI fragment, of 1.8 kb in size, were subcloned by means of using a combination digestion with the enzymes XhoI and BamHI. Combination restriction digestion with the enzymes XhoI and XbaI resulted in a XhoI/XbaI fragment of 6.5 kb in size, and two XhoI fragments, of 6.5 kb and 1.5 kb in size, respectively, being cloned.

Digestion with the restriction enzyme XhoI was used to subclone the insert, of approx. 17.9 kb in size, in phage clone P2 in the form of subfragments. In all, three XhoI subfragments, of 7.5 kb, 6.4 kb and 1.6 kb in length, respectively, were cloned. Four SacI fragments, of 4.8 kb, 3 kb, 2 kb and 1.8 kb in size, respectively, were additionally subcloned by digesting with the restriction enzyme SacI.

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The insert, of approx. 13.5 kb in size, in phage clone P3 was subcloned by digesting with the restriction enzymes SacI and/or XhoI. Six SacI subfragments, of 3.2 kb, 2 kb, 0.9 kb, 0.8 kb, 0.65 kb and 0.5 kb in length, respectively, and two XhoI subfragments, of 6.5 kb and 4.3 kb in length, respectively, were obtained in this connection.

The insert, of approx. 13.2 kb in size, in phage clone P5 was subcloned by digesting with the restriction enzymes SacI and/or XhoI. In all, SacI fragments of 6.5 kb, 3.3 kb, 3.2 kb, 0.8 kb and 0.3 kb in size, and XhoI fragmente of 7 kb and 3.2 kb in size, were subcloned.

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In order to clone the hTC genomic sequence region located 3' of phage clone P17 and 5' of phage clone P2, 3 genomic walkings were carried out using the Clontech GenomeWalker<sup>TM</sup> kits (catalogue number K1803-1) and various combinations of primers. In a final volume of 50 µl, 10 pmol of dNTP mix were added to 1 µl of human GenomeWalker Library HDL (from Clontech), and a PCR reaction was carried out in 1xKlen Taq PCR reaction buffer and 1xAdvantage Klen Taq polymerase mix (from Clontech). 10 pmol of an internal gene-specific primer, and 10 pmol of the adaptor primer AP1 (5'-GTAATACGACTCACTATAGGGC-3'; from Clontech) were added as primers. The PCR was carried out in 3 steps as a touchdown PCR. First of all, denaturation was carried out at 94°C for 20 sec, and the primers were then annealed, and the DNA chain extended, at 72°C for 4 min, over 7 cycles. There then followed 37 cycles in which the DNA was denaturated at 94°C for 20 sec but the subsequent primer extension took place at 67°C for 4 min. In conclusion, there followed a chain extension at 67°C for 4 min. After this first PCR, the PCR product was diluted 1:50. One µl of this dilution was used in a second nested PCR together with 10 pmol of dNTP mix in 1xKlen Taq PCR reaction buffer and 1xAdvantage Klen Taq polymerase mix and also 10 pmol of a nested gene-specific primer and 10 pmol of the nested Marathon Adaptor primers AP2 (5'-ACTATAGGGCACGCGTGGT-3'; from Clontech). The **PCR** conditions corresponded to the parameters which were selected in the first PCR. As the sole exception, only 5 cycles rather than 7 cycles were selected in the first PCR step and only 24 cycles, instead of 37 cycles, were run in the second PCR step. The products of this nested genomic walking PCR were cloned into the TA Cloning Vector pCRII from InVitrogen.

In the first genomic walking, the gene-specific primer C3K2-GSP1 (5'-GACGTGGCTCTTGAAGGCCTTG-3') and the nested gene-specific primer C3K2-GSP2 (5'-GCCTTCTGGACCACGGCATACC-3') were used, together with the HDL library 4, and a PCR fragment of 1639 bp in length was obtained. In the second genomic walking, a PCR fragment of 685 bp in length was amplified from the HDL library 4 using the gene-specific primer C3F2 (5'-CGTAGTTGAGCACGCTGAACAGTG-3') and the nested gene-specific primer C3F (5'-CCTTCACCCTCGAGGTGAGACGCT-3. The third genomic walking mixture, using the gene-specific primer DEL5-GSP1 (5'-GGTGGATGTGACGGCGCGTACG-3') and the nested gene-specific primer C5K-GSP1 (5'-GGTATGCCGTGGTCCAGAAGGC-3'), led to a 924 bp PCR fragments being cloned from the HDL library 1. In all, 2100 bp of the genomic hTC region located 3' of phage clone P17 were identified using this genomic walking method (see Fig. 7).

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The subcloned fragments, and the genomic walking products, were sequenced in single-stranded form. The Lasergene Biocomputing Software (DNASTAR Inc. Madison, Wisconsin, USA) was used to identify overlapping regions and form contigs. In all, 2 large contigs were assembled from the sequences collected from phage clones P12, P17, P2, P3 and P5, and also the sequence data from the genomic walking. Contig 1 consists of sequence data from phage clones P12 and P17 and the sequence data from the genomic walking. Contig 2 was put together from the sequences from phage clones P2, P3 and P5. Overlapping phage clone regions are shown diagrammaticaly in Fig. 7. The sequence data from the 2 contigs are shown below. The ATG start codon in contig 1 is underlined. The TGA stop codon is underlined in contig 2.

## Contig1:

			GGCTACGGTG					
5			AAAAAAAAA					
5			CAAGAGGAAT					
			AATGAAGAAA					
			ACCCACGGTA					
			AGAAAAGCCA					
10			TGAGGTCAGG					
10			GCTGGGCATG					
			CAGGAGGTGG					
			CTGTCTCAAG AAAAGCAAGA					
			TGAAACTGAA					
15			TTGACAAACC					
13			AGAGACATTA					
			ATAAATTGAA	•				
			AGAAATCCAA					
			AGAGAAGCCC					
20			CCTACTCAAA					
			CTGATTCCAA					
			GGCCAATATC					
			CCTTCGAAAG					
			AAATCAATCA					
25	TATGATTATT	TCACTTTATG	CAGAAAAAGC	ATTTGATAAA	ATTCTGCACC	CTTCATGATA	AAAACCCTCA	1610
	AAAAACCAGG	TATACAAGAA	ACATACAGGC	CAGGCACAGT	GGCTCACACC	TGCGATCCCA	GCACTCTGGG	1680
	AGGCCAAGGT	GGGATGATTG	CTTGGGCCCA	GGAGTTTGAG	ACTAGCCTGG	GCAACAAAAT	GAGACCTGGT	1750
			AAATTAGCCA					
20			TAAGCCTAGG					
30			AGACCCCACT					
			AGGAGGTGGA					
			ATAAAAGCCC					
			GAAAATGACA					
35			GATAAGAGAA					
33			ATGATCTTAT GATACAAAAT					
			CAAAAAAGCA					
			AAACTATAAA					
			TTGGAAGAAT					
40			TAAAATACTA					
			CCCAGAATAG					
			TATACTACAA					
			GAACAGAATA					
4.5	TTTTTGACAA	AGGTGCCAAG	AACATACTTT	GGGGAAAAGA	TAATCTCTTC	AATAAATGGT	GCTGGAGGAA	2940
45			TAACAATACT					
	GGATGAAAGG	CTTAAATCTA	AAACCTCAAA	CTTTGCAACT	ACTAAAAGAA	AACACCGGAG	AAACTCTCCA	3080
			ACTTCTTGAG					
			AAAAAGCTTC					
50			TTTGCAAACT					
50			AAAACACCTA					
			ACAAATGGCA					
			ACTATGAGAG					
			GAGGATGTGG AGTTTGAAAG					
55			CAAAAAAGGG					
55			CCAAGGTTTG					
			AATGGAGTAC					
			AGTATGTTAA					
			AAAATTAAAA					
60			GAGTCAACAA					
			GAAAGGATAA					
			GTATCAAAAT					
			GGCACGGTGG					
. <del>-</del>	GTGGATCACC	TGAGGTCAGG	AGTTTGAAAC	CAGTCTGGCC	ACCATGATGA	AACCCTGTCT	CTACTAAAGA	4340
65	TACAAAAATT	AGCCAGGCGT	GGTGGCACAT	ACCTGTAGTC	CCAACTACTC	AGGAGGCTGA	GACAGGAGAA	4410
	TTGCTTGAAC	CTGGGAGGCG	GAGGTTGCAG	TGAGCCGAGA	TCATGCCACT	GCACTGCAGC	CTGGGTGACA	4480
			ACAAAAACAA					
			AGAAGTTAAA					
70			GGGTTTCTAG					
70			GTTACTGTTG					
			TAAAGAGGCA					
			TAATTACAGA					
			TGCTTTTTT					
75			ATCCTGAAAC TGTGGACCTG					
	PAMOOUGGAMO	CHOOLGGCIC	DIJJADDIOI	AGCCACTICA	ATCTTCAAGG	0101010000	MAGACCCAGG	7110

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	TGCAAGGCAG	AGGCCTGATG	ACCCGAGGAC	AGGAAAGCTC	GGATGGGAAG	GGGCGATGAG	AAGCCTGCCT	5180
	CCTTCCTCAC	CAGCGCATGA	A CHCCCCCTTA	TTTTACCCTTTT	CCNNNCNTTC	CTCTCCATAC	CATCTCCAAA	E 2 E 0
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• ^	TGGCTGGGGG	CGGACAGCGA	CGGCGGGATT	CAAAGACTTA	ATTCCATGAG	TAAATTCAAC	CTTTCCACAT	5/40
10	CCGAATGGAT	TTGGATTTTA	ΤΥΤΑΔΤΑΤΤ	ΤΤΟΤΤΔΔΔΤΤ	ΤΓΑΤΓΑΑΑΤΑ	ACATTCAGGA	CTCCACAAAT	5810
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2.5								
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	GGAACCCGGA	GGCTGTGCCA	TCTTTGCCAT	GCCCGAGTGT	CCTGGGCAGG	ATAATGCTCT	AGAGATGCCC	///0
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		GTGAGCCACC						
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65		GTCTTCTGGG						
$\sigma_{\mathcal{I}}$								
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		TCTACTGCTG						
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	GCCTGGACCC							10/20
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	ACAGAGTGCC GCGCCTGGCT	GGGGCCCAGG CCATTTCCCA	GTCAAGGCCG CCCTTTCTCG	TTGTGGCTGG ACGGGACCGC	TGTGAGGCGC CCCGGTGGGT	CCGGTGCGCG GATTAACAGA	GCCAGCAGGA TTTGGGGTGG	10500
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			CCGCGTCTAC					
			TCCGGACCTG					
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			TCCGGGCCCT					
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			CCCGAGGCCT					
			GGGCGTGGGG					
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. •		TGACTTCTTG					•	26414
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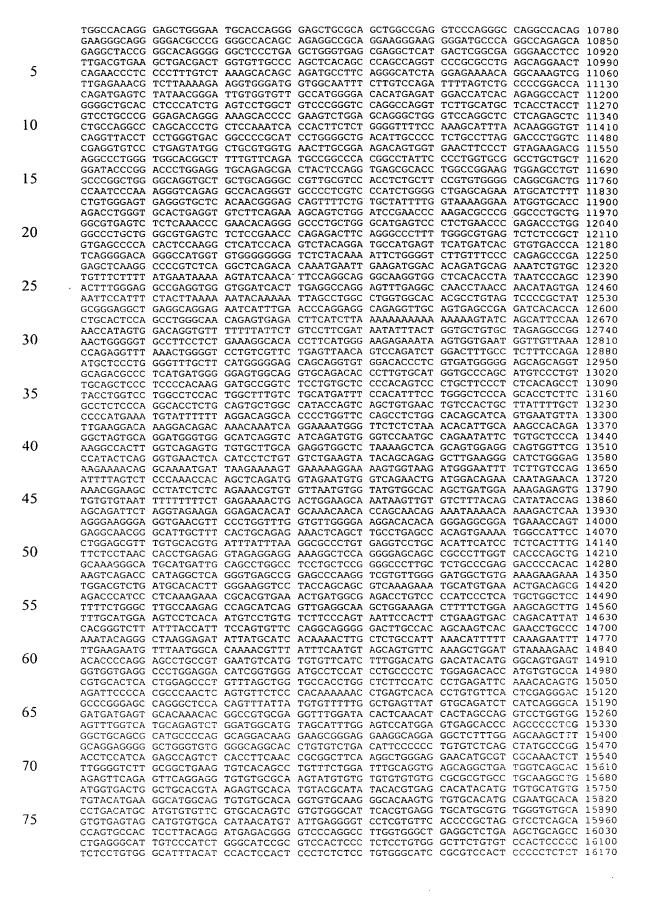
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### Contig 2:

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		GTTTGGCGTG						
1.0		CAAGTAGCTG						
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	A TOTAL CAMPAN	AACTAATCAC	TAATCCTATC	$\Lambda C C \Lambda \Lambda T T \Lambda T \Lambda$	מידית ידי די מידי מידי מ	AACTATAATT	AGAAATATTA	5000
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Le A 32 805-Fign Countries



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	CTGAACAGTA	GATGGGAGAT	CAGATGCCCG	GAGGATTTGG	GGTCTCAGCA	AAGAGGGCCG	AGGTGGGTGC	16940
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	CACCTGTGCT	CTGGGCATGG	CTGTGCTCCT	GGAACGTTCC	CTGTCCTGGC	TGGTCAGGGG	GTGCCCCTGC	17080
	CAAGAATCGA	CAACTTTATC	ACAGAGGGAA	GGGCCAATCT	GTGGAGGCCA	CAGGGCCAGC	TTCTGCCTGG	17150
15							GGCCCGGGCC	
IJ								
	TCCACCTCAA	CAGGCCTCCC	GAGCCACTGG	GAGCTGAATG	CCAGGAGGCC	GAAGCCCTCG	CCCCATGAGG	17290
	GCTGAGAAGG	AGTGTGAGCA	TTTGTGTTTAC	CCAGGGCCGA	GGCTGCGCGA	ATTACCETEC	ACACTTGATG	17360
							TACAAGGTCG	
	TACCATGAAA	ATGGTTTTTA	ACCCGAGTGC	TTGCGCCTTC	ATGCTCTGGC	AGGGAGGGCA	GAGCCACAGC	17500
20								
20							GGTGCGTCCG	
	GCTCAGACCG	CCCTCCTCTC	TGCCTTCTCT	CTCTGCCTCA	AATCTTCCCT	CGTTTGCATC	TCCCTGACGC	17640
	GTGCCTGGGC	CCTCGTGCAA	CCTCCTTCAC	TCCTTTCCGG	AAACCCTTGG	GGTGTGCTGG	ATACAGGTGC	17710
							GGGCCTCCTT	
	GGGCCATGAT	GAGGTCAGAG	GAGTTTTCCC	AGGTGAAAAC	TCCTGGGAAA	CTCCCAGGGC	CATGTGACCT	17850
25							GCTCCGAGGA	
23								
	GCTCCCGTAG	AGGGCCTGGG	CTCAGGGCAG	GGCGGCTGAG	TTTCCCCACC	CATGTGGGGA	CCCTTGGGTA	17990
	GTCGCTTGAT	TEGETAGECE	TGAGGAGGCC	GAGATGCGAT	GGGCCACGGG	CCGTTTCCAA	ACACAGAGTC	18060
							TGGGCCCCGA	
	TTTCACGGCA	GCCAGGCTGC	AGTGGGCGAG	GCTGTGGTGG	TCCACGTGGC	GCTGGGGGCG	GGGTCTGATT	18200
30							GGTGGACGCC	
2.0								
	CCGACCTCTA	GCAGGTGGCT	ATTTCTCCCT	TTGGAAGAGA	GCCCCTCACC	CATGCTAGGT	GTTTCCCTCC	18340
	TGGGTCAGGA	GOGTGGCCGT	GTGGCAACCC	CGGGACCTTA	GGCTTATTTA	TTTGTTTAAA	AACATTCTGG	18410
							AGGCTGAAAC	
	CGGGGTGCTG	GCTTGACTGG	TGTGATCTCA	GGTCATTCCA	GAAGTGGCTC	AGGAAGTCAG	TGAGACCAGG	18550
35	TACATGGGGG	CCTCACCCAC	TEGETEAGAT	GAGGTACACG	GGGGGCTCAG	GCAGTGGGTG	AGGCCAGGTA	18620
55								
							ACCAGGTACA	
	CGGGGGCTCT	GATCACACGC	ACATATGAGC	ACATGTGCAC	ATGTGCTGTT	TCATGGTAGC	CAGGTCTGTG	.18760
							CGCGGTGGCT	
							TTTAAGACCA	
4()	GCCTGAGCAA	CATAGTAGAA	CCCCATCTCT	ATGAAAAATA	AAAACAAAAA	TTAGCTGAAC	ATGGTGGTGT	18970
							GGAAGCTGCA	
	GTGAGCTGAG	ATTGCACCAC	TGTACTGCAG	CCTGGGTGAC	AGAGTGAGAG	CCCATCTCAA	СААСААСАЛА	19110
	GAAGACTGAC	AAATGCAGTT	TCTTGGAAAG	AAACATTTAG	TAGGAACTTA	ACCTACACAC	AGAAGCCAAG	19180
4.5							GGGTTTATGC	
45	ACCACAGGGG	CGGGTGGCTC	AGAAGGGATG	CGCAGGACGT	TGATATACGA	TGACATCAAG	GTTGTCTGAC	19320
	GENGGGCNGG	<b>ΛΤΤ</b> Ο ΛΤΟ ΛΤΛ	ACTACCTCCT	CCTACACAAC	CAACAATGGA	TABACTCCAA	ACCTTAGAGG	19390
	CCTTCCCGGA	ACAGGGGCTA	ATCAGAAGCC	AGCATGGGGG	GCTGGCATCC	AGGATGGAGC	TGCTTCAGCC	19460
	TCCACATGCG	TGTTCATACA	GATGGTGCAC	AGAAACGCAG	TGTACCTGTG	CACACACAGA	CACGCAGCTA	19530
							CCCATGAGGA	
50								
50	AACCCATGCA	TGTGCATTCA	TGCACGCACA	CAGGCACCGG	TGGGCCCATG	CCCACACCCA	CGAGCACCGT	19670
	CTGATTAGGA	GGCCTTTCCT	CTGACGCTGT	CCGCCATCCT	CTCAGGTTTC	ACGCATGTGT	GCTGCAGCTC	19740
							GCCTCCCTCT	
	GCTACTCCAT	CCTGAAAGCC	AAGAACGCAG	GTATGTGCAG	GTGCCTGGCC	TCAGTGGCAG	CAGTGCCTGC	19880
	CTGCTGGTGT	TAGTGTGTCA	GGAGACTGAG	TGAATCTGGG	CTTAGGAAGT	TCTTACCCCT	TTTCGCATCA	19950
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55							GAGCACCTGA	
	TGGAAGGGAC	AGGAGCTGTC	TGGGAGCTGC	CATCCTTCCC	ACCTTGCTCT	GCCTGGGGAA	GCGCTGGGGG	20090
							CCTGTGGTGG	
							TAGGAGGAGG	
	CCAGGCCCAG	GCTACCCCAC	CCCTCTCAGG	AGCAGAGGCC	GCGTATCACC	ACGACAGAGC	CCCGCGCCGT	20300
60							TCTGATCCGT	
00								
	CTGAAATTCA	AGCCATGTCG	AACCTGCGGT	CCTGAGCTTA	ACAGCTTCTA	CTTTCTGTTC	TTTCTGTGTT	20440
	GTGGAAATTT	CACCTGGAGA	AGCCGAAGAA	AACATTTCTG	TOGTGACTOO	TGCGGTGCTT	GGGTCGGGAC	20510
							GGAGGGGAGC	
	TGGGCTGGGC	CTGTGACTCC	TCAGCCTCTG	TTTTCCCCCA	GGGATGTCGC	TGGGGGCCAA	GGGCGCCGCC	20650
65	CCCCCCCCCC	CCTCCCCCCC	CCTCC TTCC	CTGTGCCACC	ש א כבר א ייייר בייי	CCTCAACCTC	ACTCGACACC	20720
55	0000010100	COLCUMBE	CGIGCWGIGG	O TO TOCCACC	ANGUNITUUI	COLOMMOCIG	managagaga	20720
	GTGTCACCTA	CGTGCCACTC	CTGGGGTCAC	TCAGGACAGG	CAAGTGTGGG	TGGAGGCCAG	TGCGGGCCCC	20790
	ACCTGCCCAG	GGGTCATCCT	TGARCGCCCT	GTGTGGGGGG	AGCAGCCTCA	GATGCTGCTG	AAGTGCAGAC	20860
	CCCCCCCC	CTCTCCC		0101000000	NOCCOUNT TOT	Character	TOCTOTOTO	20030
							TGGTGTCCCC	
	AGGCCACGGA	GCCTGGCAGG	GTCCCCAACT	TCTTGAACCC	CTGCTTCCCA	TCTCAGGGGC	GATGGCTCCC	21000
70							TGCCCTGAGC	
7.0								
	TCCTGGGGTC	CTGAGCAAGT	TCTCTCCCCC	CCCCGCCGCT	CCAGCGTCAC	TGGGCTGCCT	GTCTGCTCGC	21140
							AGGCCCTGCC	
	1 GUUUUUUU	CCCACACGTC	CIMUDAGGGT	TOCHOCHTCC	CHULILIGG	CICIICIGGA	ACGGAGTCTG	21200
	ATTTTGGCCC	CGCAGCCCAG	ACGCAGCTGA	GTCGGAAGCT	CCCGGGGACG	ACGCTGACTG	CCCTGGAGGC	21350
75							CCACAGCCAG	
, ,								
	GCCGAGAGCA	GACACCAGCA	GCCCTGTCAC	GCCGGGCTCT	ACGTCCCAGG	GAGGGAGGGG	CGGCCCACAC	21490
	CCAGGCCCGC	ACCGCTGGGA	GTCTGAGGCC	TGAGTGAGTG	TTTGGCCGAG	GCCTGCATGT	CCGGCTGAAG	21560
	CCTCACTCTC	COCCTC	CTCSCCCCC	CTCCACCCIA	COCCTCACTC	TOCACCACAC	CTGCCGTCTT	21630
	UC - CAGTGTC	COCCIONGCC	CroabCoaGT	GICCAGCCAA	COGC 1 GAG 1'G	CCAGCACAC	CTGCCGTCTT	21000

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CACTICCCAC AGGGTGGCG CTGGGTCCA CCCCAGGGC AGGTTTTCCT CACCAGGAGGC CCGGCTTCA 21700 CCCCCCCCC CATCCCAGGTG GAGACCCTG GAGACGTTG TTCACCCCCTC CCTTCCTCCT 21770 CCACCCCCCC CATCCAGGTG GAGACCCTG GAGACGTTG GGGACCTTG GGAATTTGGA GTGTGCCCTG GTGTGGCGTG TACACAGGGC AGGACCCTG AGAGGACCTG GGGACTCTG GGAATTTGGA GGGGGGGGC 21910 GTGTGGCGTG TACACAGGGC AGGACCCTG ACTGGATGG GGGTCCCTGT GGGTAAATT GGGGGGGGGC GAGACGCCAGGC CTGGAGGCC TTATACACTG GGGCCATGGGGGGT 19190 GTGCACTGCA TAGACACCAC ATTATCAGAAGC TTTGAATACC AATTCATGTT TGAAAAAAA TCTCATGTTT GGCGCCATGGCC TGGGTGCA TTTACAGGAAC TTATACAGGAA TTGAGACACCAC GGCCCATGGCC 22120 CCTGGCTGGG CCTGGAAGAC ATTACAGAACCAT CTATACAGCA GAGGGAAGGA GGGTGGGGGA TAGACACATG 22120 GACCCCCCAC CCTGGAAGAC ATAACACTAA GTCCAGGCCC GAGGGGCAAGGA GAGGAAGGA GGGGTAGGGGA TAGACACATC GCCCTCCACC TGCAGCCGGG ACGGGGATG AGGCCAGC CAGGGTGCCA GGGAAGAGGA GGGAAGGGGGA TGACACATC TCTGGGTGGC CAGGGTGGGA GGGAAGACGA GGGAAGCCTG GGGCCCACC C22400 GCCTCCACC TGCAGCCGGG GATCGTGC CAGGGTGGCA GGGAAGAGGA GGGAACCTG CTCCCCCCAC CTGCAGCCGCC CAGGGTGCCA GGGAAGACACACATC CTCAGGTGCAAC CTCCACCCACC CTCCACCCACC CTCCACCCACC		CACTTCCCCA	CACCCTCCCC	CTCCCCTCCA	CCCCACCCC	ACCTTTTCCT	CACCACCACC	CCCCCTTCCA	21700
CACCCCCAC CATCAGGTG GAGACCCTG GGAGACTTG GGATTTGGA GTARACAAGA 21840 GTGTGGCCTG TACACAGGGG AGGACCCTCC ACCTGGATGG GGGTCCCCTGT GGGTCAAATT GGGGGGAGGT 21910 GTGCACTGCA TAGACACCAC TGATACCCACCAC TATACGGAAGC TTGAGACCCAC TGATACCCACCAC TGATACCACACAC TTATACACATT TACAGAAGC TTGAGACCCAC ATACACCACCAC TAGACCCACCAC TGATACCACACAT TACAGAAGC TTGAGACCCAC ATACACCACCAC TGATACCCACAC TAGACCACCAC CTGGGAGGAC TACACACCACC CCCGGGAGGACCC CCCCGGCGCCC CCCGGGAGGACCCC CCCGGGAGGACCCC CCCCCACC CCCGGAGGACCCC CCCGGGAGGACCCC CCCCCACC CCCGGGAGACCC CCCCCACC CCCGGGAGACCC CCCCCACC CCCGGGAGACCC CCCGCCCCCC CCCGGGGCCCCC CCCCCCCC									
GTGTGCCCTG TACACAGGGG AGGACCTGC ACCTGGATGG GGGTCCCTGT GGGTCAAATT GGGGGAGGT 21910  CTGTGCGAGG TAAAATACTGA AATTATTGAG TTTTTCAGTT TGAAAAAAAA TCTGTTT GAATCTAAT 21980 GCCCATGGC TGGCTGTGCA TTTACGAAT TACAGAAGCC TGTGAGTGAA CGGGTTGGTG GTCACTGCGG 22050 GCCCTGGCTGGG CTGGCTGTGCA TTTACGAAT TACAGAGCCC TGTAGTGCAA CGGGTTGGTG GTCACTGCGG 22050 GGGCCCCCCC CTGGCAGGT TTCTGATGCT GTGAGGCAGG AGGGAAGGG GGGTCAGTGC GCGCCCATGG 22190 GGGCCCCCCC CCTGGAAGAC ATAAACAATAA GTCCAGGCCC GAAGGCACC AGGGCAGCCC AGGGCCCCAC CTGGAAGAC ATAACAATAA GTCACGCCC GAAGGCACC AGGGCACC CTGGAGGCAC AGGGCACCC GGGGTGCTG GGGGGCCCCC CTGCAGCCCG GGGATGATGG GGGGGCCCC CTCCCCCT 22300 GGGCTCCCCAC TGCAGCCGTG GATCCGGCC GGGAAATG GGGAACCTG GGGGCCCC CTCCCCCT 22300 GGCTCCCCAC TGCAGCCGTG GATCCGGATG TGCTCCCCTG GGGAAATG GGGAACCTG CTCTCCCCCT 22300 GGCTCCCCAC TGCAGCCGTG GATCCGCACC CTCTATAAAA TCCAGCATCC TCTCTCCCCT 22400 GCCTCCCACC TGCAGCCGTG GAGCCCTCC CTCATAAAA TCCAGCATTC CTCTCCTCA AGACTAGCCCAC 22410 TCAGCTTGAA AGTCACATTC CGCCTCTCCC ATCCCCTA AGACTAGCCCC AGGCTTCCTCA AGACTAGCACCC CTCTATAAAA TCCAGCATC CTCTCTCAAAAA TCCAGCATC CTCTCTCAAAAA TCCAGCATC CTCTCTCCAC AGACCACCAC CTCATCCCCTA AGACTAGCACC AGGATCCCAC AGACCACCAC CTCATCCCCAC AGACCACCAC CTCATCCCCCAC CTCACCCCCAC CACCAAACAGC AGCACCACCACCACCACCACCACCACCACCACCACCACCA									
GUTGEGGGAG TAGAACACCA TGATAFCAGAT TACAGAGAGC TGATAGAGAGAGC COTGAGTGGAG 20150 GCCCATGGCC TGGCTGTGCA TTACGGAAG TCATAGAGAGC TGATGAGTGAG GGGGGTGGT GTCATGGGG 22150 CCTGGGTGGG CUTGGGAGGT TTCTGAGTGCT GTGAGCAGG AGGGGAAGGA GGGGAAGGAG GGCCCATGGG 22120 CCTGGCTGGG CUTGGAGGAG TTCTGAGTGCT GTGAGGCAGG AGGGAAGGA GGGCAAGGAGG GGCCCACG CUTGGAGGAG AGGGCCACG CUTGGAGGCC CUTGGAGGAC AGGGCAGAG AGGGAAGAG GGGGAAGATG GGGGCCACC CUTGAGAGA GGGGAGAGAG GGGAAGATG GGGGCCACC CUTGCCCCT 22400 GCGCCCACC TGCAGCCTG GATCGGATG TCTGAGTGC GAGGAAGATG GGGAACATC GTGAGAGACCC GCTCCCCCC CACC TGGGGCCCAC CTCCCCCT 22400 GCGCCCACC TGCAGCCTG GATCGGATG TGCTTCCCTG GTGCACATC CTCTATAAAA TCCAGGATTC CTCTGGCCCC CACC TGCGCCCC CACC TGGGGGGCCAC CTCCTCCCCT 22400 GCGCTCCACC TGCAGCCAC CCTCTTCCC TTCTCTATAAAA TCCAGGATTC CTCTCCAGGACATC CTCTATACAAA TCCAGGATTC CTCTCCTGA ACCCCCCAA 22540 TCAGGTTGAA GTGCACATTC CCCCTTGGCC CATCCCTTTAT AGAGTAGACC AGGATTCCTGA ACCCCCCAA 22540 TCCCCTTTATC ATCTCCCAG GGCGGGTGC AGCCTCCCAG CCTCTCCCCTT CCTCCTCTA ACCCTCCACC TCCATCTCCCAGCT CACCATTCACC ATCCCCCAGC TCCACCTTCCCAGCT CACCATTCACC ATCCCCCAGC TCCACCTCCCAGC CACCACCAC 22840 AGCCCCCTCCT CAGAAGATTGC ATCCCCTATC CATCTCCCAGT TCCATCTCTT TCCCCAGTC TCCATCTCCC ACCACCCCCAC 22840 AGCCCCCCCC CAGAAGATTGC CAGCGAGGAGGAGGAGGAGGAGC CACCTCTGAGCAC CAGAGAACGAA AGCCCCCCCAC CAGAAGATCGA AGCCCCCCAC CAGAACATCA CACCAAAACGAA AGGGCGGGGC CAAGAGGAACCC CAGAGGAGACC CACCAGAACGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAAACCGAACCGAACCCACCCACCCCACCCCCACCCCCACCCCCACCCCCACCCC									
GTGCATGCA TAGACACCAC TGTATGCAAT TACAGAAGCC TGTGAGTGAC CGGGGTGGTG GTCATGCGG 22050  GCCCATGGC TGGCTGGCA TITACGAGAA CTATACAGGTA ATGCGGGTTG TGGGTCAGTGC GGGCCCATGG 22120  CCTGGGTGGG CCTGGAAGCA TATACGAGTA GTCAGGCAGG AGGGAAGGG GGTAGGGGA TAGACAGTGG 22190  GAGCCCCAC CCTGGAAGCA ATACACATAA GTCAGGCCC GAAGGCACA GGGCACCCAC TGGGGGCCACC 22300  GGGGTGATGG GGGGATGATGG AGGCCTGGC CAGGGTGGCA GGGAAGCACC TGGGTGGCA 22330  GGGGTGATGG GGGGGGGGG TTCTGGTGTGC GGGGAAGATG GGGCCCCACC TGGGGCCCAC 22400  GCCTCCACC TGCAGCCGTG GATCCGGATG TGCTTCCCTG GTGACACCTC TCTGGGCCAC 22470  GGAGGTGGGG GGCAGGGCA TGACACCATC CTGTATAAAAA TCCAGGATTC CTCTCTCTCAA ACCCCCCACAC  TCAGGTTGAA AGTCACATTC CGCCTTGTCTCTC CATTCTCTTA AGAGTTCTCA TCCTCCTGA ACCCCCACAC  TCCTCTATAC ATCTCCCACT CAGCGTGGC ACCACAGAGG CTTCAGGCTG GGGGTGGA TGCTCTCCACACCC  CTCTCTATAC ATCTCCCACT CTACTCTCTC ATCCTCTTTAC CAGCTTGACACCCC TCCATCCCCACT  TCTCCCAGTC TACCTCCCAG GGGGGGTGG ACCACAGGAGG CTTCAGGGTG GGGCTGGGA TGCTCTCTCACC ATCCTCTCTAC ATCCCCCAGT TACCTCCCAG TCACCACCTC TCATCTCTC ATCCCCAGT CTCATCCCCAGT TCCCCAGT TACCTCCCAG TCACCACCACT TCACTCCCAG TCACCACACCT TCACTCCCAG TCACCACCACC CCAAAACCGA AGGCCCCCACC CAGAACCCAC GGGGGGGCC CAGAGGGGGCC CAGAGGGAGC CAGCAGGGACC CAGACACCAG GGGGGGCC CAGAGGGGGCC CAGAGGGGCAC CAGACACCAC CCAAAACCGA AGGCCCTCCC CAGAGATGC CAGAGAACCCA CAGAACCGA AGGCCCTCGC TGAGGGGGCC CAGAGGACCC CAGACCACCACCACCACCACCACCACCACACCACCACCAC	5								
GCCCATGGCC TGGCTGCA TTTACGGAAG TCTATGGCAG ATTGGGTTT TGGTCAGTGC GGGCCCATGG 22190  GAGCCCCAC CCTGGAAGAC ATACAGTAA GTCCAGGCCC GAAGGGCACG AGGATGGAG GGGTAGGGGA TTTGGCGGGGC CAGGCCCAGC C22500  GGCGTGATGG GGGATGATGG AGGGCCTGGC CAGGGTGGCA GGGATGATGG GGGCCCAGC CGGGGTGCAGC CAGGGATGATGG GGGCCCAGC CAGGGATGATGG GGCCCCAGC CAGGGATGATGG GGGCCCAGC CAGGGATGATGG GGCCCCAGC CAGCAGCAGCAGC CAGGATGATGG GGGGATGATGG GGGGCCCAGC CAGCACCAGC CAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGC	5								
CCTGGCTGGG CCTGGAAGGA TATACAGTAG GTCAGGCCC GAGGGGAGGA GGGTTAGGGA TAGACAGTGG 22190  GAGCCCCCAC CCTGGAAGACA TATACAGTAG GTCAGGCCC AGAGGGAGACA GGGATCATG GGGGCCCAGC 22260  GGGTGATGG GGGGGGTGG CCTGGGTGCC CAGGGTGGCA GGGATGATG GGGCCCAGC TGGGGTGGCA 222400  GCCTCCACC TGCAGCCGT GATCGGATG TGCTTCCTG GGATAGTG GGGAAGATT GCCTCTCCCCC CTCCTCCCCC 22400  GCCTCCACC TGCAGCCGT GATCGGATG TGCTTCCTG GGATCACACCT CTTGGGCCAT CAGCTTTCAT 22470  GGAGGTGATG GGGGGGTGT GACACCATC CTTTATAAAA TCCAGGATTC CTCTCTCAA CGCCCCCAC 225400  TCCCCAGT TACATCCCAT CCCCTCTGGC CATTCCTTAT CAGAGTATC CTCCTCTCAA CGCCCCCAC 225400  TCCCCAGT TCATCTCTC ACACCTCTCC CATCCTCTTAT CAGAGTAGCAC AGGATTCTGA TCTCTCTAAACCCCAC TCCTCTCTCA TCCCCTCTTAT CATCCCCAGT CTCATCTCTC ACCCCTCTTAT CTCTCCTCATA CTCCCCAGT CTCATCTCTC TCCTCTTAT CATCCCCAGT CTCATCTCTC TCCTCTTAT CTCTCTCTATA TCCTCTTAT CTCTCCTCTAT CTCTCCAGGCA CAACCCAG AGGAACCCA GGGACCTCC AGAGGTTGC TACCTCCCAG GCCGGGTCC AGCCTCGAACCCA CAGACCCAC CAGACCACACACCAC CAGACCACACACA									
10 GAGCCCCAC CCTGGAAGAC ATAACAGTAA GTCCAGGCCC GAAGGGACG AGGATGATG GGGGCCCAGC 22230 GGGGTGATGG GGGATGATGG AGGGCCTGGC CAGGTGGCAGC GGGATGATGG GGGCCCAGC TGGGGTGCCAGC 22330 GGGGTGATGG GGGGGCATG TCTGGGTGCC GGGAGGATGATGG GGATGATGG GGCCCAGC TGGGGTGCCAC 22340 GCCTCCCACC TGCAGCCGTG GATCGGATG TGCTTCCCTG GTGCACATC TCTGGGCCAT CAGCTTTCAT 22470 GGAGGTGGGG GGCAGGGGCA TGCACACCAT CTGTATAAAA TCCAGGATTC TCTCTCCTGA AGGCCCAAC 22540 TCAGGTTGAA AGCTCACATT CGCTCTGGC CATTCTCTTA AGAGTAGCCA AGGATTCTAT CACCTCTTAT CATCTCCAGGTT CACCTCTTA ACGCCCACA 22540 TCCTCTTATC ATCTCCCAGT CTCATCTCTC ATCCTCTTA CATCTCCAGGTT CATCTCTCTA CATCTCCCAGT CTCATCTCTTA TCCTCAGGTT CATCTCTCA CATCTCTACC ATCCTCTACC ATCCTCTACC ATCCTCTACC ATCCTCTACC ATCCTCTTA TCCTCAGGTT CATCCTGTCA CACCACACACACACACACACACACACACACACACAC									
TTGGGCGGC GGGATGATGG AGGGCCTGGC CAGGGTGGCA GGGATGATGG GGGCCCCAGC TGGGGTGCA 22330 GGGGTCATGG GGGGGGCGTG TCTGGGTGGC GGGAAGATC GGGAAGATC TCTGGGCCAC CTCCTCCCCT 22400 GCCTCCCACC TGCAGCCGTG GATCCGGATG TGCTTCCCTG GTGCACATC TCTGGGCCAT CAGCTTTCAT 22470 GGAGGTGGGG GGCGGGGGA TGACACCATC CTGTATAAAA TCCAGGATTC CTCCTGA ACGCCCAAC 22540 TCAGGTTGAA AGTCACATTC CGCCTCTGGC CATTCTCTTA AGAGTAGACA AGGATTCTGA TCTCTGAGAGC 22610 TCTCCTGTATC ATCTCCCAGT CTCATCTCTCA ACACAGGAGG CTTCAGGGTG GGGCTGGTGA TGCTCTCTCA 22680 TCTCCCAGTC TCATCTGCA TCCTCTTATC ATCTCCCAGT CTCATCTTCAT TATCTCCTAAC TCTCCTCTATA CTCTCCCAGG CACACCAGGAGG CATCCTGAGA TGCCCCACAC CACACAGGAGG CATCCAGGACT TACCTCCCAG GCGGTGCC AGGGCCGCAG GGGCCCAGG CATCCTTGAC TTCCCCAGG GGAAGCAGG AGACCAGGAGGAG CATCCTTGAC TCCCCAGGC 22890 GAAGGAACTG GAAGGATTGC AGAGAACAGG AGGGCGGGC CAGGACCTCC CAGAAGCCAG AGCCCCCCC CAGAAGCCAG AGCCCCCCCC									
GGGGTGATGG GGGGGGCTG TCTGGGTGGC GGGGAAGATG GGGAAGCTG GCTGGGCCC CTCTCCCCT 22400 GCTCCCACC TGGAGCCTGGATG CGCTTCCCTG GGAGGTGGGG GGCAGGGCA TGACACATC CTGTATAAAA TCCAGGATTC CTCTGGCCCAC C22540 TCAGGTTGAA AGTCACATTC CGCCTCTGGC CATTCTCTTA AGAGGTAGCC AGGCTTCCAC 22540 TCCTCTTATC ATCTCCCAGT CACTCTCTCAC CACTCTCTCA AGGCTCCCAC 22540 TCCTCTTATC ATCTCCCAGT CACTCTCTCA TCCTCTTAT AGAGGTAGCC AGGCTTGGA TGCTCTCTA 22680 TCCTCTTATC TCATCTCCACT CTCATCTCTC ATCCTCTTAT CATCTCCCAG TCTCATCTGA TGCTCTCTA 22680 TCCTCAGGCT TCATCTCTCA TCCTCTTCC ATCCTCTTAT CATCTCCCAG TCTCATCTGA TCCTCTTAT 22750 TCTCCCAGACT TACCTCCCAG GGCGGGTGC AGGCTGGCA TGCAGCTCGA CATACAGTGG CATACAGTG GAAGGATTGC AGAGACAGA AGGCGCGGT CAGAGGCAGA CATACACTCCCAGT CATCCAGACT TACCTCCAGAC CATCCAGACT AGCCCTCC CAGAACTGG CATACAGCAGA CAGACAGA AGCCCTCC CAGAACTGG CATACAGCAG AGGCCTCAC AGAAGCAGA AGGCCCTCC CAGAACTGGA CATACACTCC CAGAACTGGA CATACACTC CAGAACTGGA CAGAGCCTAC AGCCCTACT CAGAGGACGA CAGCCTTCC CAGAGCTGC CTGGTGGGC CAGACCGA CCGAGACCT CAGAGCAGA CAGCCTACT CAGAGGACGA AGCCCTACT TCTGAGAGA CAGACACGA GCCTTATACT 23170 TGCTGCTAGATG CACCTCTGGTC CCTGGTGCC CCGGGGGC CTTATGGCCA CCTGTGACAG AGCCCTACT CAGACCTAC CACGACTCAC CACGACTCACACACTC CAGACCTAC CAGACCTCA CACGACCTCA CAGACCTAC CAGACCTAC CAGACCTCA CACGACTCACACACTC CAGACCTAC CAGACCTAC CAGACCTAC CAGACCTCA CACGACCTCA CAGACCTCA CACGACCTCA CAGACCTCA CAGACCTCA CAGACCTCA CAGACCCAC CAGACCCCA CAGACCCAC CAGACCCCA CAGACCCAC CAGACCCAC CAGACCCAC CAGACCCAC CAGACCCCA CAGACCCCA CAGACCCCA	10								
GCCTCCCACC TGGGGGCAT GACCACATC CTGTATAAAA TCCAGGATTC CTCCTCGA ACGCCCCACA 22540 GGAGGTGGGG GGAGGGGGCA TGGCACATC CTGTATAAAA TCCAGGATTC CTCCTCCTGA ACGCCCCACA 22540 TCAGGTTGAA AGTCACATC CGCCTCTGC CATTCTCTTA AGAGTAGAC AGGATTCTGA TCTCTCAAGG 22610 TCCTCTTATC ACGCCCAGT GAGGGTGGTG ACACACACT CTCATCTTA CATCTCCAGT TCCTCTTATC ATCCTCTTATC TCCTCCTATTC TCCTCCTATTC TCCTCCTATTC TCCTCCTATTC TCCTCCAGT TCCTCTTATC ATCCTCTTAT CATCTCTCAGT TCCTCTTATC TCCTCCAGT TCCTCTATC TCCTCCAGT TCCTCTCAGG C22690 CACCAGCACT TACCTCCCAG GCGGGTGCC AGGCTCGCAG CTGAGACTCTG CTTCATCTGT CTCCTCTTA 22750 GAAGGACTG GAGGATTGC CTGGGCCA AGGCGCGGGTC TGAGAGGACA CATACGTCCT TCCTCAGGCA 22890 CAGCACCCCC CAGAAGTTGG CTTGGGCCA AGGAGACAG GGGCCTGCAG GGACCTCTGGG GATGAGCTC CTGGGAGACACA AGCCACACACACA GCCCCTCTC CAGAAGTTGG CTTCAGGCAACACAG AGGCCCCTCAGAACACA GGGCCCTGGGGGC CTAATGGTAC GCCCGACACT TCCAGGAGACA GGCCCTGGGGGC CTAATGGTAC GCCGGACCACACCAC	10								
15									
TCAGGTTGAN AGTCACATTC CGCCTCTGGC CATTCTCTA ACAGTTGACC AGGATTCTGA TCTCTCAAGG 22610 GTGGGTAGGG TGGGGCAGTG GAGGTGTGG ACACAGGAGG CTTCAGGTG GGGCTGGTGA TCTCTCTCTA 22750 TCTCCCAGTC TCACTCTTAC TCTCTTTAC ATCTCCTATT CATCTCCCAG TCTCATCTGT CTTCCTTTAT 22750 CATCCAGACT TACCTCCCAG GGCGGGTGCC AGGCTCGCAG TGCATCTCTTAT TCCATGTTCT 22890 GAAGGACTT CAGAGGTTGC AGAGAACAG AGGGCGGCT TGGAGCTGGA CATACGTCT TCCTCAGGCA 22890 AGCCCCTCCT CAGAAGTTGG CTTGGGCCA AGGGAGCGGC TAGAGGGACG CATACGTCT CTCCAGGCA 22890 ACCCCCTCCT CAGAAGTTGG CTTGGGCCA AGGAACCAG AGGGCGGTGC GAGAGGCAG GGCCCTTGGG GTGAAGAACACAG AGCCACAGACACAG AGGGCGGTC TACTGAGGCA CAGAGACCAG AGCCAGAACCAG AGCCACAGACCAG AGCCACAGACAG									
15 GTGGGTAGGG TGGGGCAGTG GAGGGTTGG ACACAGGAGG CTTCAGGGTG GGCTTGGTCA TCCTCTTAC 22750 TCCCCAGTC TCATCTCTC ATCCTCTATC ATCCTCTATA CATCTCCCAG TCTCATCTGT CTCCCCTTA 22750 TCTCCCAGTC TCATCTCCAG GCCGGGTGC AGCGCTCGCAG TGCAGCTGTA CTCCCTCTTA 22750 CATCCAGACT TACCTCCCAG GGCGGTGC AGGGCCGCAG TGCAGCTGTA CTCCCTCTTA 22820 CATCCAGACT TACCTCCCAG GGCGGCTGC AGGGCGCTC TACAGCTCT CTCCCAGGCA 22890 AGCCCATCT GAGAAGTTGC CAGAACACGA GGGCCCTGCG TGAGTGGCT CAGAGGCCT CAGAGGCCT CAGAGGCCTC CAGAGGCCT CAGAGGCCC CAGAGACCGA GGCCCCAGC CAGAGCCTC CAGAGCCTTC 23030 CAGCAGGTCC CTGGTGGGGC CTTATGGTAT GGCCGGGTC TACTGAGTGC ACCTTGGAC ACCTTGGAC 22960 TTGGTGCTC TCAGAGAAT TCTGAGTTAC GCGCGGTCT TACTGATGCA GCCCTTGACA GCCTCAGTC TCTGCTGCAG CCCGCGCGC CCCCTGCT TAGAGTAC ACCTTGAGCA GCCTTCTC 23100 TTGGTGTGCT TCAGAGAACA GCGAGCCTAAT GTGTATGGTC GCCCAGAGCCT CACAGACCTT 23240 GCCCTAGGGG CGCCTTTGCC CTGCAAACC GAGACCCTAAT GTGTATGGTG GGCCCAAGTC CACAGACTT 23240 GCCCTGGGGG CGCCTTTGCC CTGCAAACC GAAGCGCAC CCCCGGGCC GCCCTGGGCG ACGACCTCAA 23380 GCGAGAGGGT GGACAGAAC GGGCGGGAC TTCCCCAGGAC CACAGCCCCCAA 23310 GCCCAGGGT TTTTTGTTGA ATTTTACTC AGGATTACTT TATTTTTTT CTAGAGAACA GTCCCCAGAG CACAGCCCCC AAGACCTCAA 23590 AAAAGGTATT TGCTTTGATA TGCCTTAACT CACTAACCAC CTACTTTATT TGCCAGCC CACCAGCT 23730 GCTGTAGCCC CACAACCCCA GGCCCATGT TAAAAACCACT ATCTTATATT TATTTATTA ATTAGAGATG GTCCTCAGAA AACCCCAC ACGACCTCAA 23590 AAAAGGTATT TGCTTTGAA AATTTTACTC AGGATTACTT TATTTATTT TTTTTTTTATT ATTTAGACATC GACACACCCCA GCCTCAGGTG ACCCCCAGGC CACAGCCCCA CCCCAGCCC CACACACCCAC AGGCCCAC GCCTCCAGCC CACACACCCAC AGCCCAC ACGACCTCC AGCCCAC CCCCACACCC ACCCACACCCAC ACCCACACCAC									
TCCTCCAGTC TCATCTCTCA ATCCTCTATA CATCTCCAGT CTCATCTCAG TCTCATCTCA	1.5								
TCTCCCAGTC TCACTCTCA TCCTCTACC AGCGGGTGC AGGCTCGAG CTCATCTCTT ATCCTCAGCA 22890  AGCCAGAACTG GAAGGATTGC AGGAGACAGG AGGCTGCAG TGGAGCTGGA CATACGTCCT TCCTCAGGCA 22990  AGCCCTCCT CAGAAGTTGC CTGGGCCA CAGAACCGA GGGCCTCCAGGGGCC CACTCTTGGG GTGAAGAAAC 22960  CAGCAGGTCC CTGGTGGGGCC CTTATGGTAT GGCCGGGTC TACTGAGTGC TCAGAGCCTTC 23030  CAGCAGGTC CTGGTGGGGCC CTTATGGTAT GGCCGGGTC TACTGAGTGC ACCTTGGACTA GGGCTCTTACT TCAGAGAACCAGA GGCCTCATT TTTTAGATGCA GCCCGAGCTTATT GGCCGGGCCT TACTGAGTGC ACCTTGGACTA GGCCTCAGACTA TTTTTTTTTT	15								
20 GARGCACT TACCTCCAG GEGGGGGGCC AGGCCGCAG TGGAGCTGCA CATACGTCCT TCCTCAGGCA 22890 AGCCCTCCT CAGAACTTGC CTTGGGCCAC ACGAACCGA GGCCCTCTGC TGAGTGGCTC CAGAGCCTTC 23030 TTTGAGTGCA GCCCGACCT GCCTGGTTC GGCGGGGGCC TACTGAGTCA CACTTGGACA GGCCTTCTGG GTGAAGACCC 23100 TTTGAGTGCA GCCCGGACCT GCCTGGTTC GGGGTGGGGC TTACTGACTGC ACCTTGGACA GGCCTTCTGC 23100 TGCTGCTGCT TCAGAGAATG TCTGAGTGCC CTGGATGCG CGTCATTTTAT 23170 TGCTGCTGCT TCAGAGAATG TCTGAGTGCC CCGGACTG GGGTCCTTATGGCCA ACCTTGGACA GGCCTTCTGC 23100 GGCCTGGGGG CGCCTTTGCC CTGGAGCCC CCGTATAGGCG GGCCCAGATAG GGCCATATT 23170 GGCCTGGGGG CGCCTTGCC CTGCAAACTG GAAGGCAGG GCCCCGGGCC CCGTGGAGG GCCCTGACC C32310 GGCCTGGGGG CGCCTTTGCC CTGCAAACTG GAAGGCAGGC GCCCCGGGCC CCGTGGGGGG ACGACCTCAA 23380 GTGAACACACA ACCACAGGT CAGGCCATTG TTCAGGCACC CACCAGGAC CACCTGGGTT 23450 TGAATCACAG ACCACAGGT CAGGCCATTG TTCAGGCTATC CACTCTTCAC AAAGCTCCA ATCCTCTGTT 23520 CTCCGGGTGT TTTTTGTATA TGCTTTAGTA TTCCTAGGCA CACCTCTATC CACTAACTA TAATTTTTT TTTTATATT ATTAGAAGATA GGGTCTAACT ACCTCTACC ACCTAGCTAC CACTAACTACT ATATTTTTT TTTTTATTATT AGACCCCCA GGCTCAAGTG ATCCTCCTATC CACTAACGAC CTACTTTATT TGTCTTTTATT ATATACAGAGTA GTGTCTAACT CACTAAGCAC CTACTTTATT TATTTATTATT ATTAGAGACT GGGCCAAGCCC CTACTAAGTA ATCCTCCTAC CACTAAGCAC CTACTTTATT TATTTATTATT ATAGAGACT AGGCCACCC CTACTAAGGTC CACCAGCCCC CTACCTAGCT CACTAAGCAC CTACTTTATT TATTTATTATT ATAGAGACC CAAGCCCCA GGCCAAGCCC CACCTGGCC CACCTGGCC CTACAGCCC CTACAGCCC CAAGCCCCA GGCCAAGCCCCA CACCAGCCCCA CACCAGCCCCCA CACCAGCCCCA CA									
20 AGGCATCTC CAGAAGTTGC CTTGGGCAC ACGAAACCGA AGGCGCCTC CAGAGGGCCC CAGAGGCCTC CAGAAGTTGC CTTGGGCAC ACGAAACCGA ACCTTGGCTC CAGAAGTTGC CTTGGGCAC CTTGGTGTGC GCCTGGTGTC GGCCTGCGTT CAGAGGTGCC CTTGGTGTC CAGAGAGCTT CZ3030 TTTGAGTGCA GCCCGGACCTT CTTGGTGTC CAGAGAGCTT CTTGGTGTC CAGAGAGCTC CAGAGCTC CAGAGCTC CAGAGCTC CAGAGCTCC CAGAGATGC CTTGGTAGAGAG GCCCAAGT GTTATGGCA CACTTGGACA GGCCTATTAT Z3170 TGCTGCTGCTGC CAGAGAGTG CCTGGAGCCC CCGTATAGGG GCCCAAGT GTTATGGCA CACGAGCCTC ACAGAGCTGC ACAGAGCTGC ACAGAGCAGCACAA GCCACACAGG CCCTGGAGCC CCGTGAGGGA CACCTGGAGACC CAGAGCCTCA CAGAGCCTAA GGCCTAAGT GTTATGGGAGA ACAACACAGG CCCCTGGAGCC CCGTGAGGAG ACCACCAGAGC CCGGAGGCCC CCGTGGGGGA CACCTGGGT Z3450 TGAAACACACA ACAACAGGT CAGAGCCATA TTCAGCAGAG CAGAGCGCC CCGTGGGGCA ACCACCAGAG CCCCAGGGC CCGTGGGGA CACCTGGGT Z3450 TGAAACACACA ACAACAGGT CAGAGCCATA TTCAGCATAT CAGAGCACTCAA AAATTTATT ATTATATATT ATTAGAGATG GTGTCTAACT ACACCACA GCCCAAGACCCCA GCCCTAGAGCAC CACCTGGGT CAGAGCCATA ACACCACA AGACCCCAA GCCCAAGAGC CACCTGGAGT CACACACACACACACACACACACACACACACACACACA									
ACCCCTCCT CAGAAGTTGG CTTGGGCCA ACGAACCGA GGGCCCTGCG TAGTGGCTC CAGAGCCTTC 23030 CAGCAGGTCC CTGGTGGGGC CTTATGGATA GGCCGGGTCC TACTGAGTGC ACCTTGGACA GGGCTTCTGG 23100 TTTGAGTGCA GCCCGGACGT GCCTGGTTC GGGGTGGGGC CTTATGGCCA CTGGATATGG CGTCATTTAT 23170 TGCTGCTGCT TCAGAGAATG TCTGAGTGC CAGACCCTATT GTGTATGGTG GCCCAAGTC CACAGACTGT 23240 GTCGTAAATG CACTCTGGTG CCTGGAAACTG GAAGGGAGGG GCCCTGAGGA AGGACGGCC CCGAGCCC CCGAACCC CCGAGCCC CCGAGCCC CCGTATAGGA GCCCCGGGCG CGCTTGCCACCC CTGCAAACTG GAAGGGAGGG GCCCTGGGCG AGCACCTCAA 23380 GTGAGAGGTT GGACACACGGC CAGGCCATT TTCCACGGAG CAGAGCCCCC CGTGGGCG ACGACCTCAA 23380 CTCCGGGTG TTTTTGTTGA AATTTTACTC AGGACTACC CATCTTCTTAC AAAACCTCCAC ATTCCTGTT 23520 CTCCGGGTG TTTTTAGACATG GTGTCAACCCCA ACCACACCTT TCACACACACCCAC CACCTGGTT 23550 AAAAGGTATT TGCTTTGATA TGGCTTACC ACCTAAGCAC CACCTAGACCCAC ATTCCTGTT 23520 CTGTCAGCCC CAAACCCCCA GGCCCAACCC CACCACACCCCAC CACCTAGACCAC CACCTAGACCAC CACCTAGACCAC CACCACACCA									
CAGCAGGTCC CTGGTGGGGC CTTATGGTAT GGCCGGGTCC TACTGAGCA CCTTGGACA GGCCTTCTGG 23100 TTTGAGTGCA GCCCGGACGT GCCTGGTGTC GGGGTGGGGG CTTATGGCCA CTGGATATG CGTCATTAT 23170 TGCTGCTGCTT TCAGAGAATG TCTGAGTGAC CGAGCCTAAT GGTGATAGGT GGCCCAGCTC CACAGACTG 23240 GTCGTAAATG CACTCTGGTG CCTGGAGCCC CCGAGCCTAAT GGTGATAGGT GGCCCAGGGG CCTGGGGG CCCCGGGGG CCCGTGGGGG CCCGTGGGGG CCCGTGGGGG CCCGGGGG CCCGTGGGGG AGGACCTCAA 23380 GTGAGAGGTT GGACACAAC GGGCCATTG TTCCAGGAG CCCCGGGGC CCCTGGGGG CCCTGGGGG CCCCCGGGGC CCCCCGGGGC CCCCCGGGGC CCCCCGACCTC AAAAGACACAACAACAACAACAACAACAACAACAACAACA	30								
TTTGAGTGCA GCCCGGACGT GCCTGGTGT GGGGTGGGG CTTATGGCCA CTGGATATGG CGCTATTAT 23170 TGCTGGTGCT TCAGAGAATG TCTGAGTGAC CGAGCCTAAT GTGTATGGT GCCCAAGTC CACAGACTGT 23240 GGCCTAGAATG GCCCTGGAGCC CGGTATAGGA GCTGTGAGGA AGGAGGGGCT CTGGCAGCC 23310 GGCAGGGGG GCCCTGGGGG GCCCTGGGGG GCCCGGGGG CCCGTGGGGG AGGACCTCAA 23380 GTGAACACAG ACCAACAGGT CAGGCCATT TTCAGCTAC CATCTTCTAC AAAGCTCCAG ATCCTGGTT 23520 CTCCGGGGT TTTTTGTA ATTTATACC AGGATATCT CATCTTCTAC AAAAGCTCCAG ATCCTGTT 23520 AAAAGGTATT TGGCTTAACT CACTAACCAC CATCTTCTAC AAAAGCTCCAG ATCCTGTT 23520 AAAAAGGTATT ATTTATATA ATTTATACC AGGATTACT TATTTATTAT AGACCCTTAA 23590 AAAAGGTATT ATTAGAGATG GTGCTAACT CACTAAGCAC CTACTTTATT TGTCTGTTT TATTATATA 23660 TATTATTATTA ATTAGAGATG GTGCTCACC TGGCCAC GCCCCAGGC CCCAGGTGC GCCCAGGTG TCTTAGGCAC CACTAGCAC CTACTTTAGT TATTATTAT 23660 TGTGAGCCAC CACCCCCA GGCTCAACGG CACCTCACTTC TATTATTATT ATTATATAT 23660 TGTGAGCCAC CACCCCCA GGCTCAACGG CACCTCACCT	20								
TGCTGCTGCT TCAGAGAATG TCTGAGTGAC CGAGCCTAAT GTGTATGGTG GGCCCAAGTC CACAGACTGT 23240 GTCGTAAATG CACTCTGGTG CCTGGAGCC CCGGTATAGGA GCTGTAGAGA AGGAGGGGCT CTTGCCAGCC 23310 GGCCTGGGGG CCCCGGGCG CCCTGGGGCG CCTGCAGCC 23310 GGCCTGAGACCA ACCAACAGCT CAAGGACCTA AAAAAAAAAA									
GTCGTAAATG CACTCTGGTG CCTGGAGCCC CCGTATAGGA GCTGTGAGGA AGGAGGGCT CTTGGCAGCC 23310 GGCCTGGGGG CGCCTTTGCC CTGCAAACTG GAAGGGAGCG CCCCGGGCG CGTGGGGCG ACGACCTCAA 23380 GTGAGAGGTT GCACCAGACA GGCCGGGCA TTCCCAGGGA CAGAGGCCC TGCTCAGGCA CACCTGGGTT 23450 TGAATCACAG ACCAACAGGT CAGGCCATTG TTCCCAGGGA CAGAGGCCC TGCTCAGGCA CACCTGGGTT TTTTTCTTGA AATTTTACT CAGGATTACTT ATATTTTTTTG CTAAAGTATT AGACCTTAA CACTAAGCAC CTACTTGTATT TTTTCTGTATA TGCTTTGATA TGCCTTAACT CACTAAGCAC CTACTTGTATT TTTTCTGTTTTT TATTTATTT ATTAGAGATG GTCCTCACC CTACCTTACT TATTTTTTTT TATTTATTT									
GGCCTGGGGG CGCCTTTGCC CTGCAAACTG GAAGGAGCCG CCCGGGCG CCGTGGGCGG ACGACCTCAA 23380 GTGAGAGGTT GGACAGAACA GGGCGGGGAC TTCCCAAGGAC CAAGGCCCG TGCTCAGGCA CACCTGGGTT 23450 CTCAGGGTG TTTTTGTTGA AATTTACTC AGGATTACTT CATCTTTAC AAAGCTCCAG ATTCCCTGTTT 23520 CTCCGGGTGT TTTTTGTAA AATTTTACTC AGGATTACTT ATATTTTTTG CTAAAGTATT AGACCCCTAA 23590 AAAAGGTATT TGCTTTGATA TGGCTTAACT CACTAAGCAC CTACTTTATT TGTCTGTTTT TATTTATTAT 23660 GCTGTAGCCG CAAACCCCCA GGCTCAAAGTG ATCCCTCGGC CTCAGGTTC CAGAGTGCTC GGCATTACAGCAC CTACTTTATT TGTCTGTTTT TATTTATTAT 23660 GCTGTAGCCG CAAACCCCCA GGCTCAAAGTG ATCCCTCGGC CTCAGCTTC CAGAGTGCTC GGCATTACAGC 23800 CTGTCATCCC AGTAGCTCC TGGCACTTTT AAAAACCACT ATGTAAGGTC AGGTCCAGTG GCTCATACAG 23800 CTGTCATCCC AGTAGTTTGG GAAGCCCAGT CTCTACAAA AAATGCAAAAA AGTTAATCCGG GCTTCCACAC 23870 CTGCAACATAG GGAGACCCCA TCCCTACAAA AAATGCAAAAA AGTTAATCCGG GCGTGGGTC CAGCATCTC 24010 AGGCCAAGCT GCCCTGCC TGAGTGGGA GCACGAGGTT GTCTGAGGCC AGGAGGAGC CAGCAGGAGG AGAAGGAAAA AAAAAAAA									
GTGAGAGGTT GGACAGACA GGGCGGGGAC TTCCCAGGAG CAGAGGCCGC TGCTCAGGCA CACCTGGGTT 23450 TGAATCACAG ACCAACAGGT CAGGCCATTG TTCAGCTATC CATCTTCTAC AAAGCTCCAG ATTCCTGTTT 23520 AAAAGGTATT TGCTTTGATA TGGCTTAACT CACCAAGCAC CTACTTTATT TGTCTGTTTT TATTTATTA 23560 TATTATTATTATT ATTAGAGATG GTGCTTAACT CACCAAGCAC CTACTTTATT TGTCTGTTTT TATTTATTAT 23660 TGTGAGCCAC TGCCCTTGCC TGGCACTTTT TATTAGACACAC CTACTTCCACCAG GTCAAGGCCC CAGAGGACCCCA GCCTCAGCTCC CAGAGTGCTC CAGAGTGCTC CAGAGTCCCAG GTCAAGGCC CAGAGGACCCA TGCCCTTACAAA AAAAACCACA AGGTCAAGTC AAAAAACCACA AGGTCCAAGT GCCTCAGCTCC CAGAGTCTC CAGCTTCC CAGAGTCCCA GGTTCACACC 23870 CTGTCATCCC AGCACTCC AGCACTGGC CAGAGGATT AAAAACCACA AGGTCCAAGT GCCTCAGCTC CAGCTTCC CAGCTTCC CAGCTTCC CAGCTTCC CAGCTTCC CAGCTTCC CAGCTTCC CAGCTCC CAGCATCGCC CAGCACCAT CTCTCACAAA AAAAACAAAA AAATACAAAA AAAAAAAA	2.5								
TGAATCACAG ACCACAGGT CAGGCCATTG TTCAGCTATC CATCTTCTAC AAAGCTCCAG ATTCCTGTTT 23520 AAAAAGGTATT TGCTTTGATA TAGGCTTAACT CACTAAGCAC CTACTTTATT TGCTGTTTT AGACCCTTAA 23690 TATTATTATT ATTAGACAGT GTGTCTACT TGTCACCCAG GTGTTAGTG CAGTAGCCC CTACTTATT TGTCTGTTTT TATTATATTA	25								
30 TATATTATT TOTTGATA TOTTGATA TOTTGATCA CACTAAGCAC CACTATTATT TOTTGATCA TATATTATT TOTTGATCA TOTTGATCA TOTTGATCACT TOTTGATCACT TOTTGATCACT CACTAAGCAC CACTAGGAC CAGAGGACT CAGAGGACAC GAAACCCCCA GGCTCAAGCAC ATCCTCCAG GTTGTTAGT CAGAGGACAC GCATCATCACC CAGACTCCC CAGACTGCT CAAACTACAGACAAAAAAAAAA									
30 TATTATTATT TGCTTTGATA TGGCTTAACT CACTAAGCAC CTACTTATT TGTCTGTTT TATTTATTAT 23660 GCTGTAGCCCC CAAACCCCCA GGCTCAAGTG ATCCTCCGGC CTCAGCTTCC CAGAGTGCTAC GGCTCAAGTG CTGCCCTTGCC TGGCACTTTT AAAAACCACT ATGTAAGGTC CAGAGTGCTAC CAGAGTGCTAC CAGAGTGCTAC AGGACCCCA ACTCTCCAGCT AGGACCCCA ACTCTCACAAA AAATCCAAAA AGTTATCCGG GCTCCAGCCT CAGCATCTG CAGCACCCAGCAC CAGAGAGACAC AGGACACCCA ACGACACCAC AGCACCCAGCAC CAGCACCCAGCAC CAGCACCCAGCAC CAGCACCCAGCAC CAGCACCAC CAGCACCAC CAGCACCAC CAGCACCAC CAGCACCCAC CAGCACCAC CAGCACCCAC CAGCACCCAC CAGCACCCAC CAGCACCAC CAGCACCACACACA									
TATTATTATT ATTAGAGATG GTGTCTACTC TGTCACCCAG GTTGTTAGTG CAGTGGCACA GTCATGGCTC 23730 GCTGTAGCCG CAAACCCCCA GGCTCAAGTG ATCCTCCGGC CTCAGCTTC CAGAGTGCTG GGATTACAGG 23800 TGTGAGCCAC TGCCCTTGCC TGGCACTTTT AAAAACCACT ATGTAAGGTC AGGTCCAGTG GGTTCCACCAC 23870 CGTAACATAG GGAGACCCCA TCTCTACAAA AAATGCAAAA AGGAGTTTG GCGACTTTG AGGACCAGGT CAGGAGTTTGA GACCAGCATG 23940  35 AGTCCCAGCT GCTCGGGAGG CTGAGTGGA AAATGCAAAA AATTCCGG GCGTGGGGT CAGCATCTGT 24010 AGGAGAGAGG AGAAGGAAG AAGAAGGAGG AACCCTGTCTC AAAAAAAAAA		CTCCGGGTGT	TTTTTGTTGA	AATTTTACTC	AGGATTACTT	ATATTTTTTG	CTAAAGTATT	AGACCCTTAA	23590
GCTGTAGCCG CAAACCCCCA GGCTCAAGTG ATCCTCCGGC CTCAGCTTCC CAGAGTGCTG GGATTACAGG 23800 TGTGAGCCAC TGCCCTTGCC TGGCACTTTT AAAAACCACT ATGTAAGGTC AGGTCCAGTG GCTTCCACAC 23870 CTGTCATCCC AGTAGTTTGG GAAGCCGAGG CAGAAGGATT GTCTGAGGCC AGGAGTTTGA GACCCAGCATG 23940 GGTAACATAG GGAGACCCCA TCTCTACAAA AAATGCAAAA AGTTATCCGG GCGTGGGGTC CAGCATCTGT 24010 AGTCCCAGCT GCTCAGGAGG CTGAGTGGGA AAATGCAAAA AGTTATCCGG GCGTGGGGTC CAGCATCTGT 24010 AGGAGAGAAGG AGAAGAAGGA AAGAAGGAAG AAGAAG	20								
TGTGAGCCAC CTGTCATCCC CTGTCATCCC CTGTCATCCC CTGTCATCCC CTGTCATCCC CGGAGACCCCA TCTCTACAAA AAAACCACT CTGTCAGGCC AGGAGACCCCA TCTCTACAAA AAATGCAAAA AAATGCAAAA AGTTCTCGGGCC AGGAGACCCCA TCTCTACAAA AAATGCAAAA AGTTCTCGGGCC AGGAGCCCCA TCACACAAA AAATGCAAAA AGTTCTCCGGGCC AGGAGCCCCA TCACACAA AAAGAGAAAA AGTCCCGGGGG GGTAGCGTGGGCC AGCCTGGGCA AAAAAAAAAA	30								
CTGTCATCCC AGTAGTTTGG GAAGCCGAGG CAGAAGGATT GTCTGAGGCC AGGAGTTTGA GACCAGCATG 23940 GGTAACATAG GGAGACCCCA TCTCTACAAA AAATGCAAAA AGTTATCCGG GCGTGGGGTC CAGCATCTGT 24010 AGTCCCAGCT GCTCGGGAGG CTGAGTGGGA GGATCGGTTG AGCCCGGGAG GTCATGGCTG CAGTAGCTG CAGTGAGCTG AACGCAGCAGCATCGGTGA ACGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGC									
35 AGTACATAG GGAGACCCA TCTCTACAAA AAATGCAAAA AGTTATCCGG GCGTGGGGTC CAGCATCTGT 24010 AGTCCCAGCT GCTCGGAGG CTGAGTGGA AGCACTTG AGCCCGGAG GTCATGGCTG CAGTGAGCTG 24080 TGATTGTACC ATCGCACTCC AGCCTGGGCA ACAGAGTGAG ACCCTGTCTC AAAAAAAAAA		TGTGAGCCAC	TGCCCTTGCC	TGGCACTTTT	AAAAACCACT	ATGTAAGGTC	AGGTCCAGTG	GCTTCCACAC	23870
ACTCCCAGCT GCTCGGGAGG CTGAGTGGGA GGATCGCTTG AGCCCGGGAG GTCATGGCTG CAGTGAGCTG 24080 TGATTGTACC ATCGCACTCC AGCCTGGCA ACAGAGTGAG ACCCTGTCTC AAAAAAAAAA		CTGTCATCCC	AGTAGTTTGG	GAAGCCGAGG	CAGAAGGATT	GTCTGAGGCC	AGGAGTTTGA	GACCAGCATG	23940
TGATTGTACC ATCGCACTCC AGCCTGGGCA ACAGAGTGAG ACCCTGTCTC AAAAAAAAAA									
AAGGAGAAGG AGAAGGAAG AAGAAGGAAG AAGAAGGAAG AAGAAG	35	AGTCCCAGCT	GCTCGGGAGG	CTGAGTGGGA	GGATCGCTTG	AGCCCGGGAG	GTCATGGCTG	CAGTGAGCTG	24080
40 AAGGAGGCCT GCTAGGTGCT AGGTAGACTG TCAAATCTCA GAGCAAAATG AAAATAACAA AGTTTTAAAG 24290 ACAAAGCAGAAAAAAAAAAAAAAAAAAAAAAAAAAAAA		TGATTGTACC	ATCGCACTCC	AGCCTGGGCA	ACAGAGTGAG	ACCCTGTCTC	AAAAAAAAA	AAAAAAAAAG	24150
40 ACAAGCAGGT ATGGAGCTC TTTGGACTTC CTTAGGCCTG AACTTCATCT CAAGCAGCTT CCTTCCACAG 24360 ACAAGCAGGT ATGGAGCAGG TGAGTTCAAA GCAGAAAGGG AGGAGAAGCA GGCAAGGGT GAGGCTGTGG 24430 GTGACACCAG CCAGGACCCC TGAAAAGGGA TGGTTGTTTT CCTGCCCAG CCCCACGCTC CTGCCGGTCC 24500 TCACCTGCT GTAACCGTCG ATGGTGGC CAGGTGCCCA CCTGGGAAGG ATGCTGTGCA GGGGCCTTGC 24570 CAAACTTTGG TGGGTTTCAG AAGCCCCAGG CACTTGTGGC AGGCACAATT ACAGCCCCTC CCCAAAGATG 24640 CCCACGTCCT TCTCCTGGAA CCTGTGAATG TGTCACCCGC AAGGCACAATT ACAGCCCCTC CCCAAAGATG 24710 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGTGAGAGAG GGAGGCAGGG GAGAGTCCAG GAGAAGGAC ACTGGCCACT 24850 GCTGCCTTTG AGATGGAGGA GGGGCCCCC AGCCAAGGAA TGGGGCCAGG CGCCCACACC CGCCCCACGCCC AGCAAAGACCA 24920 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTC TCTCAGCCCA CTAAGCTGCA GGGACCTGTT 24990 ACAGCAAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGCTGA TCTCAGCCCA 25130		AAGGAGAAGG	AGAAGAGAAG	AAGAAGGAAG	AAGGAAAGAG	AAGAAGAAGG	AAGAAGGAAG	AAAGAAGGAG	24220
40 ACAAGCGTGT ATGGAGCGAG TGAGTTCAAA GCAGAAAGGG AGGAGAAGCA GGCAAGGGTG GAGGCTGTGG 24430 GTGACACCAG CCAGGACCCC TGAAAGGGAG TGGTTGTTTT CCTGGCCCACGCTC CTGCCGGTCC 24500 TGCACCTGCT GTAACCGTCG ATGGTGTGCCCA CCTGGGAAGG ATGCTGTGCA GGGGCTTGC 24570 CCAAACTTTGG TGGGTTTCAG AAGCCCCAGG CACTTGTGGC AGGCACAATT ACAGCCCCTC CCCAAAGATG 24640 CCCACGTCCT TCTCCTGGAA CCTCTGAATG TGTCACCCGC AAGGCAGAGG CTGGTGAAGG CTGCAGGTGG 24710 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGGGAGAGG GGGGCCAGG GAGAGTCAGA GAGGGGACGT GAGAAGGAC ACTGGCCACT 24850 GCTGGCTTTG AGATGGAGGA GGGGTCCCC AGCCAAGGAA TGGGGGCAGG CGCTCCATGC TGGAAAAGCA 24920 ACCAATCCTC CCCGGTCCTG AGGGCACAGG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGGCTGAC TCTCAGCCCA 25130									
GTGACACCAG CCAGGACCC TGAAAGGGAG TGGTTGTTTT CCTGCCTCAG CCCCACGCTC CTGCCGGTCC 24500 TGCACCTGCT GTAACCGTCG ATGTTGGTGC CAGGTGCCCA CCTGGGAAGG ATGCTTGTGCA GGGGGCTTGC 24570 CAAACTTTGG TGGGTTTCAG AAGCCCCAGG CACTTGTGGC AGGCACAATT ACAGCCCCTC CCCAAAGATG 24640 CCCACGTCCT TCTCCTGGAA CCTGTGAATG TGTCACCCCG AAGGCAGAGG CTGGTGAAGG CTGCAGGTGG 24710 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGTGAGAGAG GGGGGCAGGG GAGAGTCAGA GAGAGAGACC ACTGGCCACAAG 24780 GCTGGCTTTG AGATGAGAGAG GGGGGTCCCC AGCCAAGGAA TGGGGGCAGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTTGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		GGAAAGAAAA	ACCCCAGCTC	TTTGGACTTC	CTTAGGCCTG	AACTTCATCT	CAAGCAGCTT	CCTTCCACAG	24360
TGCACCTGCT GTAACCGTCG ATGTTGGTGC CAGGTGCCCA CCTGGGAAGG ATGCTGTGCA GGGGGCTTGC 24570 CAAACTTTGG TGGGTTTCAG AAGCCCCAGG CACTTGTGGC AGGCACAATT ACAGCCCCT CCCAAAAGATG 24640 CCCACGTCCT TCTCCTGGAA CCTGTGAATG TGTCACCCGC AAGGCAGAGG CTGGTGAAGG CTGCAGGTGG 24710 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGTGAGAGAG GGAGGCAGGG GAGAGTCAGA GAGGGGACGT GAGAAGGACC ACTGGCCACT 24850 GCTGGCTTTG AGATGGAGGA GGGGGTCCCC AGCCAAGGAA TGGGGCAGGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130	40	ACAAGCGTGT	ATGGAGCGAG	TGAGTTCAAA	GCAGAAAGGG	AGGAGAAGCA	GGCAAGGGTG	GAGGCTGTGG	24430
CAAACTTTGG TGGGTTTCAG AAGCCCCAGG CACTTGTGGC AGGCACAATT ACAGCCCCTC CCCAAAGATG 24640 CCCACGTCCT TCTCCTGGAA CCTGTGAATG TGTCACCCGC AAGGCAGAGG CTGGTGAAGG CTGCAGGTGG 24710 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGTGGAGGAG GGAGGCAGGG GAGAGTCAGA GAGGGGACCT GAGAAGGACC ACTGGCCACT 24850 GCTGGCTTTG AGATGGAGGA GGGGCACCC AGCCAAGGAA TGGGGCAGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130									
45 CCCACGTCCT TCTCCTGGAA CCTGTGAATG TGTCACCCGC AAGGCAGAGG CTGGTGAAGG CTGCAGGTGG 24710 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGTGAAGAGG GGAGGCAGGG GAGAGTCAGA GAGGAGAGCC ACTGGCCACT 24850 GCTGGCTTTG AGATGGAGGA GGGGGTCCCC AGCCAAGGAA TGGGGGCAGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTCTAAGAT GATGCGTTT TCTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		TGCACCTGCT	GTAACCGTCG	ATGTTGGTGC	CAGGTGCCCA	CCTGGGAAGG	ATGCTGTGCA	GGGGGCTTGC	24570
45 AATCACGGCT GCCAGTCAGC CGATCTTAAG GTCATCCTGG ATTATCTGGT GGGCCTGATA TGGCCACAAG 24780 GGTCCCTAGA AGTGAGAGAG GGAGGCAGGG GAGAGTCAGA GAGGGGACGT GAGAAGGAC ACTGGCCACT 24850 GCTGGCTTTG AGATGGAGGA GGGGGCTCCCC AGCCAAGGAA TGGGGGCAGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		CAAACTTTGG	TGGGTTTCAG	AAGCCCCAGG	CACTTGTGGC	AGGCACAATT	ACAGCCCCTC	CCCAAAGATG	24640
GGTCCCTAGA AGTGAGAGAG GGAGGCAGGG GAGAGTCAGA GAGGGGACGT GAGAAGGACC ACTGGCCACT 24850 GCTGGCTTTG AGATGGAGGA GGGGGTCCCC AGCCAAGGAA TGGGGGCAGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		CCCACGTCCT	TCTCCTGGAA	CCTGTGAATG	TGTCACCCGC	AAGGCAGAGG	CTGGTGAAGG	CTGCAGGTGG	24710
GCTGGCTTTG AGATGGAGGA GGGGGTCCCC AGCCAAGGAA TGGGGGCAGC CGCTCCATGC TGGAAAAGCA 24920 AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130	45	AATCACGGCT	GCCAGTCAGC	CGATCTTAAG	GTCATCCTGG	ATTATCTGGT	GGGCCTGATA	TGGCCACAAG	24780
AGCAATCCTC CCCGGTCCTG AGGGCACACG GCCCTGCCCA CGCCTCGATT TCAGGCCAGT GGGACCTGTT 24990 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		GGTCCCTAGA	AGTGAGAGAG	GGAGGCAGGG	GAGAGTCAGA	GAGGGGACGT	GAGAAGGACC	ACTGGCCACT	24850
50 TCAGCTTTCC GGCCTCCAGA GCTGTAAGAT GATGCGTTTG TGTTCAGCCA CTAAGCTGCA GTGATTCGTC 25060 ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		GCTGGCTTTG	AGATGGAGGA	GGGGGTCCCC	AGCCAAGGAA	TGGGGGCAGC	CGCTCCATGC	TGGAAAAGCA	24920
ACAGCAGCAA ATGGAATAGC AGTACAGGGA AATGAATACA GGGACAGTTC TCAGAGTGAC TCTCAGCCCA 25130		AGCAATCCTC	CCCGGTCCTG	AGGGCACACG	GCCCTGCCCA	CGCCTCGATT	TCAGGCCAGT	GGGACCTGTT	24990
		TCAGCTTTCC	GGCCTCCAGA	GCTGTAAGAT	GATGCGTTTG	TGTTCAGCCA	CTAAGCTGCA	GTGATTCGTC	25060
CCCCTGGG 25138	50	ACAGCAGCAA	ATGGAATAGC	AGTACAGGGA	AATGAATACA	GGGACAGTTC	TCAGAGTGAC	TCTCAGCCCA	25130
		CCCCTGGG							25138

#### Example 5

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Comparison of the above-described genomic hTC sequence and the sequence of the hTC cDNA (Fig. 6: corresponding to SEQ ID NO 2) made it possible to elucidate the exon-intron structure of the hTC gene. The genomic organization of the hTC gene is illustrated diagrammatically in Fig. 7. The coding region of the hTC gene is composed of 16 exons which vary in size between 62 bp and 1354 bp (see Table 1). Exon 1 contains the translation start codon ATG. The translation stop codon TGA and the 3'-untranslated region lie on exon 16 (Fig. 8). No possible polyadenylation signal (AATAAA) was found either in exon 16 or in the 3195 bp of the following

Le A 32 805-F

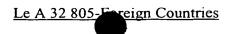
ign Countries

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3'-flanking region. The exon-intron transitions were determined on the basis of the consensus sequence

		5'-Exon				Intron							3'-Exon		
5	Pre-mRNA	A/C	A	G	G	T	A/G	A		N C	A	G	G		
	Frequency (%)	70	60	80	100	100	95	70		80	100	100	60		

and listed in Table 1. With the exception of the 5' splice site between exon 15 and intron 15, all the exon-intron transitions are in accord with the published (Shapiro and Senapathy, 1987) splice consensus sequence. The sizes of the introns are between 104 bp and 8616 bp. Since only part of intron 6 was isolated, it is not possible to determine the precise length of the hTC gene. Based on the part sequence of ~4660 bp, which was obtained from intron 6, the minimum size of the hTERT gene is 37 kb.



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Introns 1-5 and the 5' region of intron 6, are contained in contig 1:
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Intron 1: bp 11493-11596 (SEQ ID NO 4);

Intron 2: bp 12951-21566 (SEQ ID NO 5);

Intron 3: bp 21763-23851 (SEQ ID NO 6);

5 Intron 4: bp 24033-24719 (SEQ ID NO 7);

Intron 5: bp 24900-25393 (SEQ ID NO 8);

5' region of intron 6: bp 25550-26414 (SEQ ID NO 9).

The 3' region of intron 6, and introns 7-15, are located in contig 2 at the following

10 positions:

3' region of intron 6: bp 1-3782 (SEQ ID NO 10);

Intron 7: bp 3879-4858 (SEQ ID NO 11);

Intron 8: bp 4945-7429 (SEQ ID NO 12);

Intron 9: bp 7544-9527 (SEQ ID NO 13);

15 Intron 10: bp 9600-11470 (SEQ ID NO 14);

Intron 11: bp 11660-15460 (SEQ ID NO 15;

Intron 12: bp 15588-16467 (SEQ ID NO 16);

Intron 13: bp 16530-19715 (SEQ ID NO 17);

Intron 14: 19841-20621 (SEQ ID NO 18);

20 Intron 15: 20760-21295 (SEQ ID NO 19).

The 3'-untranscribed region is also located in contig 2 at position 21960-25138 (SEQ ID NO 20).

The individual sequences of the abovementioned introns are as follows:

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## Intron 1 (SEQ ID NO 4)

GTGGGCCTCCCCGGGGTCGGCGTCCGGCTGGGGTTGAGGGCGGCCGGGGGGAACCAGCGACATGCGGAGAGCAGCGCAGG CGACTCAGGGCGCTTCCCCCGCAG

## 5 Intron 2 (SEQ ID NO 5)

CTGGTCCTCCTGTCTCCATCGTCACGTGGCCACACGTGGCTTTTCGCTCAGGACGTCGAGTGGACACGGTGATCTCTGCC TCTGCTCTCCTGTCCAGTTTGCATAAACTTACGAGGTTCACCTTCACGTTTTGATGGACACGCGGTTTCCAGGCGC CGAGGCCAGAGCAGTGAACAGAGGAGGCTGGGCGCGCAGTGGAGCCGGGTTGCCGGCAATGGGGAGAAGTGTCTGGAAG CACAGACGCTCTGGCGAGGGTGCCTGCAGGTTACCTATAATCCTCTTCGCAATTTCAAGGGTGGGAATGAGAGGTGGGGA CGAGAACCCCCTCTTCCTGGGGGTGGGAGGTAAGGGTTTTGCAGGTGCACGTGGTCAGCCAATATGCAGGTTTGTGTTTA AGATTTAATTGTGTGTTGACGGCCAGGTGCGTGGCTCACGCCGGTAATCCCAGCACTTTGGGAAGCTGAGGCAGGTGGA TCACCTGAGGTCAGGAGTTTGAGACCAGCCTGACCAACATGGTGAAACCCTATCTGTACTAAAAAATACAAAAATTAGCTG GGCATGGTGGTGTGCCTGTAATCCCAGCTACTTGGGAGGCTGAGGCAGGAGAATCACTTGAACCCAGGAGGCGGAGGC CGTTGATTGTGCCAGGACAGGGTAGAGGGAGGGAGATAAGACTGTTCTCCAGCACAGATCCTGGTCCCATCTTTAGGTAT GAAGAGGCCACATGGGAGCAGAGGACAGCAGATGGCTCCACCTGCTGAGGAAGGGACAGTGTTTTGTGGGTGTTCAGGGG ATGGTGCTGCTGGGCCCTGCCGTGTCCCCACCCTGTTTTTCTGGATTTGATGTTGAGGAACCTCCGCTCCAGCCCCCTTT TGGCTCCCAGTGCTCCCAGGCCCTACCGTGGCAGCTAGAAGAAGTCCCGATTTCACCCCCTCCCCACAAACTCCCAAGAC AAAAGTCATATAACATGAGATTGGCACTCCTAACACCGTTTTCTGTGTACAGTGCAGAATTGCTAACTCGGCGGTGTTTA CAGCAGGTTGCTTGAAATGCTGCGTCTTGCGTGACTGGAAGTCCCTACCCATCGAACGGCAGCTGCCTCACACCTGCTGC GAGAGTTTGAGTTCTCTGATCAGGACTCTGCCTGTCATTGCTGTTCTCTGACTTCAGATGAGGTCACAATCTGCCCCTGG GTCACGTGTAGGGTGAGTGAGGCGCGCCCCCGGGTGTCCCTGTCCCGTGCAGCGTGATTGAGGTGTGCCCCCGGGTGT GAGGCTCTGTCCCCAGGTGTCCTTGGCGTTTGCTCACTTGAGCTTGCTCCTGAATGTTTGCTCTTTCTATAGCCACAGCT GCGCCGGTTGCCCATTGCCTGGGTAGATGGTGCAGGCGCAGTGCTGGTCCCCAAGCCTATCTTTTCTGATGCTCGGCTCT TCTTGGTCACCTCTCCGTTCCATTTTGCTACGGGGACACGGGACTGCAGGCTCTCGCCTCCCGCGTGCCAGGCACTGCAG TCTCCCAGCTTGTCTCATGCCGAGGCTGGACTCTGGGCTGCCTGTGTCTGCCACGTGTTGCTGGAGACATCCCAGAA AGGGTTCTCTGTGCCCTGAAGGAAAGCAAGTCACCCCAGCCCCTCACTTGTCCTGTTTTCTCCCAAGCTGCCCCTCTGC TTGGCCCCCTTGGGTGGGTGGCAACGCTTGTCACCTTATTCTGCGCACCTGCCGCTCATTGCTTAGGCTGGGCTCTGCCT GAGGGCCGGTGTCTCCGCCAGCCTTCGTCAGACTTCCCTCTTGGGTCTTAGTTTTGAATTTCACTGATTTACCTCTGACG TTTCTAFCTCTCCATTGTATGCTTTTCTTGGTTTATTCTTTCATTCCTTTTCTAGCTTCTTAGTTTAGTCATGCCTTTC CCTCTAAGTGCTGCCTTACCTGCACCCTGTGTTTTGATGTGAAGTAATCTCAACATCAGCCACTTTCAAGTGTTCTTAAA AATCATTTTGATATCAGTGACTTTTAAGTATTCTTTAGCTTATTCTGTGATTTCTTTGAGCAGTGAGTTATTTGAACACT GTTTATGTTCAAGATATGTAGAGTATCAAGATACGTAGAGTATTTTAAGTTATCATTTTATTATTGATTTCTAACTCAGT TGTGTAGTGGTCTGTATAATACCAATTATTTGAAGTTTGCGGAGCCTTGCTTTGTGATCTAGTGTGCATGGTTTCCAG 

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 ${\tt AAGCTTCTGTCTCCTTCTAGATGCATGAAATTCCAAGAAGGAGGCCATAGTCCCTCACCTGGGGGATGGGTCTGTTCATT}$ TCTTTTGGAGACTTCTATGTCTCTAGTAATCTAGTAATTCTTTTTTTAAATTGCTCTTAGTACTGCCACACTGGGCTTCT GAGTCTTGGTCTGTCGCCCAGGGTGAGTGCAGTGGTGTGATCACAGGTCAGTGTAACTTTTACCTTCTGGCCTGAGCCGT CCTCTCACCTCAGCCTCCTGAGTAGCTGGAACTGCAGACACGCACCGCTACACCTGGCTAATTTTTAAATTTTTTCTGGA GACAGGGTCTTGCTGTTTGCCCAGGCTGGTCTCAAACTCTTGGACTCAAGGGATCCATCTACCTCGGCTTCCCAAAGTG CTGAATTACAGGCATGAGCCACCATGTCTGGCCTAATTTTCAACACTTTTATATTCTTATAGTGTGGGTATGTCCTGTTA ACTAGAGACCCGCCTGGTGCACTCTGATTCTCCACTTGCCTGTTGCATCGTTCCCTTGTTTCTCACCACCTCTTG GGTTGCCATGTGCGTTTCCTGCCGAGTGTGTTGATCCTCTCGTTGCCTCCTGGTCACTGGGCATTTGCTTTATTTCT CTTTGCTTAGTGTTACCCCCTGATCTTTTATTGTCGTTGTTTGCTTTTGTTTATTGAGACAGTCTCACTCTGTCACCCA GGCTGGAGTGTAATGGCACAATCTCGGCTCACTGCAACCTCTGCCTCCGCTTCAAGCAGTTCTCATTCCTCAACCTCA TGAGTAGCTGGGATTACAGGCGCCCACCACCACGCCTGGCTAATTTTTGTATTTTTAGTAGAGATAGGCTTTCACCATGT TGGCCAGGCTGGTCTCAAACTCCTGACCTCAAGTGATCTGCCCGCCTTGGCCTCCCACAGTGCTGGGATTACAGGTGCAA GCCACCGTGCCCGGCATACCTTGATCTTTTAAAATGAAGTCTGAAACATTGCTACCCTTGTCCTGAGCAATAAGACCCTT AGTGTATTTTAGCTCTGGCCACCCCCAGCCTGTGTGCTGTTTTTCCCTGCTGACTTAGTTCTATCTCAGGCATCTTGACA CCCCCACAGCTAAGCATTATTAATATTGTTTTCCGTGTTGAGTGTTTCTGTAGCTTTGCCCCCGCCCTGCTTTTCCTCC TTATTGCTGGTAAACCCCAGCTTTACCTGTGCTGGCCTCCATGGCATCTAGCGACGTCCGGGGACCTCTGCTTATGATGC ACAGATGAAGATGTGGAGACTCACGAGGAGGGCGGTCATCTTGGCCCGTGAGTGTCTGGAGCACCACGTGGCCAGCGTTC CTTAGCCAGTGAGTGACAGCAACGTCCGCTCGGCCTGGGTTCAGCCTGGAAAACCCCAGGCATGTCGGGGTCTGGTGGCT CCGCGGTGTCGAGTTTGAAATCGCGCAAACCTGCGGTGTGGCGCCAGCTCTGACGGTGCTGCCTGGCGGGGGAGTGTCTG CTTCCTCCCTTCTGCTTGGGAACCAGGACAAGGATGAGGCTCCGAGCCGTTGTCGCCCAACAGGAGCATGACGTGAGCC ATGTGGATAATTTTAAAATTTCTAGGCTGGGCGCGGTGGCTCACGCCTGTAATCCCAGCACTTTGGGAGGCCAAGGCGGG TGGATCACGAGGTCAGGAGGTCGAGACCATCCTGGCCAACATGATGAAACCCCATCTGTACTAAAAACACAAAAATTAGC TTTGTCTGCGGGATCCCGTGTGTAGGTCCCGTGCGTGGCCATCTCGGCCTGGACCTGCTGGGCTTCCCATGGCCATGGCT GTTGTACCAGATGGTGCAGGTCCGGGATGAGGTCGCCAGGCCCTCAGTGAGCTGGATGTGCAGTGTCCGGATGGTGCACG GCCCTCGGTGAGCTGGAGGTATGGAGTCCGGATGATGCAGGTCCGGGGTGAGGTCGCCAGGCCCTGCTGTGAGCTGGATG TGTGGTGTCTGGATGGTGCAGGTCAGGGGTGAGGTCTCCAGGCCCTCGGTAAGCTGGAGGTATGGAGTCCGGATGATGCA AGGCCCTGCGGTGAGCTGGGTGTGCGGTGTCTGGATGGTGCAGGTCTGGAGGTCGCCAGACGGTGCCAGACCATGC GGTGAGCTGGATATGCGGTGTCCGGATGGTGCAGGTCTGGGGTGAGGTTGCCAGGCCCTGCTGTGAGTTGGATGTGGGGT GCCCTCGGTGAGCTGGATGTGCAGTGTCCAGATGGTGCAGGTCCGGGGTGAGGTCGCCAGACCCTGCGGTGAGCTGGATG TGCGGTGTCTGGATGCTGCAGGTCTGGAGTGAGGTCGCCAGGCCCTCGGTGAGCTGGATGTATGGAGTCCGGATGGTGCC GGTCCGGGGTGAGGTCGCCAGACCCTGCTGTGAGCTGGATGTGCGGTGTCTGGATGGTACAGGTCTGGAGTGAGGTCGCC AGACCCTGCTGTGAGCTGGATATGCGGTGTCCGGATGCTGCAGGTCAGGGTGAGGTCTCCAGGCCCTCGGTGAGCTGGA  $\tt GGTATGGAGTCCGGATGATGCAGGTCCGGGGTGAGGTCGCCAGGCCCTGCTGAACTGGATGTGCGGCGTCTGGATGGT$ GCAGGTCTGGGGTGTGGTCGCCAGGCCCTCGGTGAGCTGGAGGTATGGAGTCCGGATGATGCAGGTCCGGGGTGAGGTCG CCAGGCCCTGCTGTGAGCTGGATGTGCGGCGTCTGGATGGTGCAGGTCTGGGGTGTGGTCGCCAGGCCCTCGGTGAGCTG

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GAGGTATGGAGTCCGGATGATGCAGGTCCGGGGTGAGGTTGCCAGGCCCTGCTGTGAGCTGGATGTGCTGTATCCGGATG GTGCAGTCCGGGGTGAGGTCGCCAGGCCCTGCTGTGAGCTGGATGTGCTGTATCCGGATGGTGCAGGTCTGGGGTGAGGT CACCAGGCCCTGCGGTGAGCTGGTTGTGCGGTGTCCGGTTGCTGCAGGTCCGGGGTGAGTTCGCCAGGCCCTCGGTGAGC TGGATGTGCGGTGTCCCGTGTCCGGATGGTGCAGGTCCAGGGTGAGGTCGCTAGGCCCTTGGTGGGCTGGATGTGCCGT GTCCGGATGGTGCAGGTCTGGGGTGAGGTCGCCAGGCCTTTGGTGAGCTGGATGTGCGGTGTCTGCATGGTGCAGGTCTG GCTGTGAGCTGGATGTGCGGTGTCTGGATGGTGCAGGTCCGGGGTGAGGTAGCCAAGGCCTTCGGTGAGCTGGATGTGGG GTGTCCGGATGGTGCAGGTCCGGGGTGAGGTCGCCAGGCCCTGCGGTTAGCTGGATATGCGGTGTCCGGATGGTGCAGGT CCGGGGTGAGGTCACCAGGCCCTGCGGTTAGCTGGATGTGCGGTGTCTGGATGGTGCAGGTCCGGGGTGAGGTCGCCAGG CCCTGCTGTGAGCTGGATGTGCTGTATCCGGATGGTGCAGGTCCGGGGTGAGGTCGCCAGGCCCTGCAGTGAGCTGGATG TGCTGTATCCGGATGGTGCAGGTCTGGCGTGAGGTCGCCAGGCCCTGCGGTTAGCTGGATATGCGGTGTCGGATGGTGCA GGTCCGGGGTGAGGTCACCAGGCCCTGCGGTTAGCTGGATGTGCGGTGTCCGGATGGTGCAGGTCTGGGGTGAGGTCGCC AGGCCCTGCTGTGAGCTGGATGTGCTGTATCCGGATGGTGCAGGTCCGGGGTGAGGTCGCCAGGCCCTGCGGTGAGCTGG ATGTGCTGTATCCGGATGGTGCAGGTCTGGCGTGAGGTCGCCAGGCCCTGCGGTGAGCTGGATGTGCAGTGTACGGATGG CGCCAGGCCCTGCGGTGAGCTGGATGTGTGTGTCTGGATGCTGCAGGTCCGGGGTGAGTTCGCCAGGCCCTCGGTGAGC TGGATATGCGGTGTCCCGTGTCCGAATGGTGCAGGTCCAGGGTGAGGTCGCCAGGCCCTTGGTGGGCTGGATGTGCCGT GTCCGGATGCTGCAGGTCTGGGGTGAGGTCGCCAGGCCCTTGGTGAGCTGGATGTGCGGTGTCCGGATGGTGCAGGTCCG GGGTGAGGTCACCAGGCCCTCGGTGATCTGGATGTGGCATGTCCTTCTCGTTTAAG

## Intron 3 (SEQ ID NO 6)

GTACTGTATCCCCACGCCAGGCCTCTGCTTCTCGAAGTCCTGGAACACCAGCCCGGCCTCAGCATGCGCCTGTCTCCACT TGCCTGTGCTTCCCTGGCTGTGCAGCTCTGGGCTGGGAGCCAGGGGCCCCGTCACAGGCCTGGTCCAAGTGGATTCTGTG AAGCAGAAGGGATTTAAATTAGATGGAAACACTACCACTAGCCTCCTTGCCTTTCCCTGGGATGTGGGTCTGATTCTCTC TCTCTTTTTTTTTTTTTTTTGAGATGGAGTCTCACTCTGTTGCCCAGGCTGGAGTGCAGTGGCATAATCTTGGCTCACT GCAACCTCCACCTCCTGGGTTTAAGCGATTCACCAGCCTCAGCCTCCTAAGTAGCTGGGATTACAGGCACCTGCCACCAC GCCTGGCTAATTTTTGTACTTTTAGGAGAGACGGGGTTTCACCATGTTGGCCAGGCTGGTCTCGAACTCATGACCTCAGG TGATCCACCCACCTTGGCCTCCCAAAGTGCTGGGTTTACAGGCTAAGCCACCGTGCCCAGCCCCGATTCTCTTTTAATT CAGGGAGCACCTGTGCAGGAGCACCTGGGGATAGGAGAGTTCCACCATGAGCTAACTTCTAGGTGGCTGCATTTGAATG GCTGTGAGATTTTGTCTGCAATGTTCGGCTGATGAGAGTGTGAGATTGTGACAGATTCAAGCTGGATTTGCATCAGTGAG GGACGGGAGCGCTGGTCTGGGAGATGCCAGCCTGGCTGAGCCCAGGCCATGGTATTAGCTTCTCCGTGTCCCGCCCAGGC TGACTGTGGAGGGCTTTAGTCAGAAGATCAGGGCTTCCCCAGCTCCCCTGCACACTCGAGTCCCTGGGGGGGCCTTGTGAC ACCCCATGCCCCAAATCAGGATGTCTGCAGAGGGAGCTGGCAGCACCTCGTCAGAGGTAACACAGCCTCTGGGCTGGG AATGCACCTTACTTAGACTTTACACGTATTTAATGGTGTGCGACCCAACATGGTCATTTGACCAGTATTTTGGAAAGAAT TTAATTGGGGTGACCGGAAGGAGCAGACAGACGTGGTGGTCCCCAAGATGCTCCTTGTCACTACTGGGACTGTTGTTCTG CCTGGGGGGCCTTGGAGGCCCCTCCTCCTGGACAGGGTACCGTGCCTTTTCTACTCTGCTGGGCCTGCGGCCTGCGGTC AGGGCACCAGCTCCGGAGCACCCGCGGCCCCAGTGTCCACGGAGTGCCAGGCTGTCAGCCACAGATGCCCAGGTCCAGGT GTGGCCGCTCCAGCCCCGTGCCCCCATGGGTGGTTTTGGGGGAAAAGGCCAAGGGCAGAGGTGTCAGGAGACTGGTGGG CTCATGAGAGCTGATTCTGCTCCTTGGCTGAGCTGCCCTGAGCAGCCTCTCCCGCCCTCTCCATCTGAAGGGATGTGGCT CTTTCTACCTGGGGTCCTGCCTGGGGCCAGCCTTGGGCTACCCCAGTGGCTGTACCAGAGGGACAGGCATCCTGTGTGG AGGGGCATGGGTTCACGTGGCCCCAGATGCAGCCTGGGACCAGGCTCCCTGGTGCTGATGGTGGGACAGTCACCCTGGGG GTTGACCGCCGGACTGGGCGTCCCCAGGGTTGACTATAGGACCAGGTGTCCAGGTGCCCTGCAAGTAGAGGGGCTCTCAG  $\tt TGAGTCGGTGGGGGTTGCCGTTGAGCTTCCCCTTAGTCTGTTGTCTGGCTGAGCAAGCCTCCTGAGGGGGCTCT$ CTATTGCAG



#### Intron 4 (SEQ ID NO 7)

#### Intron 5 (SEQ ID NO 8)

## 5'-region intron 6 (SEQ ID NO 9)

## 3'-region intron 6 (SEQ ID NO 10)

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CAGAAGAGTTTCACGTGTGCTGATTTCCCGGCTGTTTCCTGCGTAATTGGTGTCTGCTGTTTATCGATGGCCTCCTTCCA TCTAAACAAGCATCTGAAGTTGCCGTTTTCCCTCTAAAGCAGGGATCCCGAGGCCCCTGGCTGTGGAGTGGCACCGGTCT GGGGCCTGTTAGGAACCCGGCGCACAGCGGGAGGCTAGGTGGGGTGTGGGGAGCCAGCGTTCCCGCCTGAGCCCCGCCCC TCTCAGATCAGCAGTGGCATGCGGTGCTCAGAGGCGCACACACCCTACTGAGAACTGTGCGTGAGAGGGGTCTAGATTCT GTGCTCCTTATGGGAATCTAATGCCTGATGATCTGAGGTGGAACCGTTTGCTCCCAAAACCATCCCCTTCCCCACTGCTG TCCTGTGGAAAAATCGTCTTCCACGAAACCAGTCCCTGGTACCACAATGGTTGGGGACCCTGTGCTAAAGACCTGCTTCA GCAGCCTCTCGTCAGTGTTGATATATTGGCTTTTCTGTGTTGAGTCCAGAATAATTACGGATTTCTGTGATGCTTTCCGC CGACCTCAGACCCATGGGCTATTTGTGGGCGTGTTGCCTGCTCCTGGGTTGGGAAGGGTGCAGGCCCCATGTACCTTCCT GTTACTGCCTTCCAGGTTGGTTCTCAGGGTTGAATCGTACTCGATGTGGTTTTAGCCCACGGCCCTGCCGCCAGCTCCTG GGGGCTGGGGAACATGCTGAAGCACAGAGTCACCGTGCGCGTCTTTTGATGCCTCACAAGCTCGAGGCCTCCTGTGTCCG TGTTAGTGTGTCACGTGCCTGCTCACATCCTGTCTTGGGGACGCAGGGGCTTAGCAGGTCCCGTAGTAAATGACAAGC GTGCCTGCACCTGCATCCCTGCAATCCCTCCAGCACTGGGCTGGAGAGGCCCGGGAGCTCGAGTGCCACTTGTGCCACGT GACTGTGGATGGCAGTCGGTCACGGGGGTCTGATGTGTGGTGACTGTGGATGGCGGTTGGTCACAGGGGTCTGATGTGTG GTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGGTGACTGTGG ATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTG GGGTCTGATGTGGTGACTGTGGATGGCAGTCGTGGGGTCTGATGTGTGACTGTGGATGGCGGTCGTGGGGTCTGATG TGGTGACTGTGGATGGCAGTCGTGGGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGGTGACT GTGGATGGCGGTCGTGGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGG CGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGTGATCGGTCA CAGGGGTCTGATGTGTGGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGACTGTGGATGGTGATCGGTCACAG GGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGGTGACTGTGGATGGCGGTTGGTCCCGGGGG TCTGATGTGGTGACTGTGGATGGCGATCGGTCACAGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCT GATGTGTGGTGACTGTGGATGCCGTCGTGGGGTCTGATGTGTGGTGACTGTGGATGCCGGTCGTGGGGTCTGATGTGGT GACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGGTGACTGTGGAT GGCGGTTGGTCCCGGGGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCAG TCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGATGTGATGTGATGGCGGTCGTGGGG TCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGT GGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGGTGACTGTGGATGGTGATCGGTCACAGGGGTCTGATGTGTGGT GACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGAT GGCGGTCGTGGGGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTAGGGTCTGATGTGTGGTGACTGTGGATGGCAGTCG GTCACAGGGGTCTGATGTGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGTGATGTGACTGTGGATGGCGGTCGTGG GGTCTGATGTGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGATGTGGTGACTGTGGATGGCGGTCGTGGGGTCTGAT GTGGTGACTGTGGATGGTGATCGGTCACAGGGGTCTGATGTGTGGTAGCTGCAGGTGGAGTCCCAGGTGTGTCTGTAGCT ACTTTGCGTCCTCGGCCCCCGGCCCCCGTTTCCCAAACAGAAGCTTCCCAGGCGCTCTCTGGGCTTCATCCCGCCATCG GGCTTGGCCGCAGGTCCACACGTCCTGATCGGAAGAAACAAGTGCCCAGCTCTGGCCGGGGCAGGCCACATTTGTGGCTC ATGCCCTCTCCTCTGCCGGCAG

## 40 Intron 7 (SEQ ID NO 11)

GTCTGGGCACTGCCCTGCAGGGTTGGGCACGGACTCCCAGCAGTGGGTCCTCCCCTGGGCAATCACTGGGCTCATGACCG
GACAGACTGTTGGCCCTGGGGGGCAGTGGGGGGAATGAGCTGTGATGGGGGCATGATGAGCTGTGCCTTGGCGAAATC
TGAGCTGGGCCATGCCAGGCTGCĠACAGCTGCTGCATTCAGGCACCTGCTCACGTTTGACTGCGGGCCTCTCTCCAGTT
CCGCAGTGCCTTTGTTCATGATTTGCTAAATGTCTTCTCTGCCAGTTTTGATCTTGAGGCCAAAGGAAAGGTGTCCCCCT
CCTTTAGGAGGGCAGGCCATGTTTGAGCCGTGTCCTGCCCAGCTGCCCCTCAGTGCTGAGGCCAAAGGAAAGGAAACG
TGTCCCCCTTCTTAGGAGGACACGGCCCGTGTTTGAGCCACGCCCCGCTGAGCGGGCCTCTCAGTGCTGGGTCTGTCCACGT

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#### Intron 8 (SEQ ID NO 12)

TCCCGAGGCCCGGAAACATGGCTCGGCTTGCGGCAGCCGGAGCGGAGCAGGTGCCACACGAGGCCTGGAAATGGCAAGC GGATTTTATCCGATTCTCATTCCTGTCCTGTCGTGTGACCCCCGCGAGGGCGCGGGGCTCTTCTCTCTGTGACTAGATTT CCCATCTGGAAAGTGCGGGGTTGACCGTGTAGTTTGCTCCTCTCGGGGGGGCCTGTGGTGGCCATGGGGCAGGCGGCCTGG GAGAGCTGCCGTCACACACCCCTGGGTGAGCCACACTCACGGTGGTAGAGCCACAGTGCCTGGTGCCACATCACGTCCT CACAAATTGCACATGGCAGCAGAGTGAATTTTGGCCGAGGGACACGTGTGCACATGTGTGTAAGCGGCCCCCAGGCCCAC AGAATTCGCTGACAAAGTCACCTCCCCAGAGAAGCCACCACGGGCCTCCTTCGTGGTCGTGAATTTTATTAAGATGGATC GGTGACTGTGTCTGTCCCTAGGACACGGACAGGCCCGAAGCTCTAGTCCCCATCGTGGTCCAGTTTGGCCTCTGA  $\mathsf{TCTGCTTGCGTTGACTCGCTGGCCTGGCCGGACTCCTAGAGTTGGTGCGTGTGCTTCTGTGCAAAAAGTGCAGTCCTCTT$ CTGTTGTCTGCCTGGGCTTGAGTGCAGTGGCGCGATCTCAACTCACTGCAACCTCCGCCTCCCGGGTTCCAGCATTTCTC GGATTACAGGTGTGAGCCATCACGCCCAGCCGGAAAGCCTCTTTTTAAGGTGACCACCTATAGCGCTTCCCGAAAATAAC TCGCGTGGCAGCCATGCCTTCTGTGTGCACCTTTAGGTTCCACGGGGGCTATTCTGCTCTCACTGTTTGTCTGAAAACGCA CCCTTGGCATCCTTGTTTGGAGAGTTTCTGCTTCTCGTTGGTCATGCTGAAACTAGGGGCAAGGTTGTATCCGTTGGCGC AGAGCAAGGATGTGGTCACACCTGTGGCTGGATCTGTTTCAGCCGCCCCAGTGCATGGTGAGAGTGGGGAGCAGGGATTG TTTGTTCAGAGGTCTCATCTGGTATGTTTCTGAGGTGTTTTGCCGGCTGAATGGTAGACGTGTCGTTTGTGTGTATGAGGT TCTGTGTGTGTGTGGGCTCGGTTTGAGTGTACGCATGTCCAGCACATGCCCTGCCCGTCTCTCACCTGTGTCTTCCCGC CCCAG

## Intron 9 (SEQ ID NO 13)

GTGAGGCCTCCTCTCCCCAGGGGGGCTTGGGTGGGGGTTGATTTGCTTTTGATGCATTCAGTGTTAATATTCCTGGTGC TCTGGAGACCATGACTGCTCTGTCTTGAGGAACCAGACAAGGTTGCAGCCCCTTCTTGGTATGAAGCCGCACGGGAGGGG

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GAGGGCCGCTGCCTGCATGATGAGCATGTGAATTCAACACCGAGGAAGCACCAGCTTCTGTCACGTCACCCAGGTTC  $\tt CTGGGCGCCTCTTCAGCCCATTGCCATCCCACTTGCATGGGGTCTACACCCCAAGGACGCCACACCCTAAATATCGTGCC$ ATGTGCACGACGTGCAGGTTAGTTACATATGTATACATGTGCCATGTTGGTGTGCTGCACCCATTAACTCATCATTTACA GCTCAGAGTGATGGTTTCCAGCTTCGTCCATGTCCCTACAAAGGACATGAACTCATCCTTTTTTATGACTGCATAGTATT GTGAATAGTGCCGCAATAAACATACGTGTGCATGTGTCTTTATAGCAGCATGATTTATAATCCTTTGGGTATATACCCAG TAATGGGATGGCTGGGTCAAATGGTATTTCTAGTTCTAGATCCTTGAGGAATCACCACACTGTCTTCCACAATGGTTGAA  $\tt CTAGTTTACACTCCCACCAACAGTGTAAAAGTGTTCTGGTGCTGGAGAGGATGTGGACAGCAGTTATTTTTTATGAAAA$ TAGTATCACTGAACAAGCAGACAGTTAGTGAAGGATGCGTCAGGAAGCCTGCAGGCCACACAGCCATTTCTCTCGAAGAC TCCGGGTTTTTCCTGTGCATCTTTTGAAACTCTAGCTCCAATTATAGCATGTACAGTGGATCAAGGTTCTTCTTCATTAA GGTTCAAGTTCTAGATTGAAATAAGTTTATGTAACAGAAACAAAATTTCTTGTACACACAACTTGCTCTGGGATTTGGA GGAAAGTGTCCTCGAGCTGGCGGCACACTGGTCAGCCCTCTGGGACAGGATACCTCTGGCCCATGGTCATGGGGCGCTGG ACAGCTGCCATGCTGGTAAAGGGCACCACGTGGCTCAGAGGGGGCGAGGTTCCCAGCCCCAGCTTTCTTACCGTCTTCAG GCTGATGGTAAACACTGAGTACTTATAATGAATGAGGAATTGCTGTAGCAGTTAACTGTAGAGAGCTCGTCTGTTGGAAA TCGTAGACAGATACTACGTAAAAAGTGTAAAGTTAACCTTGCTGTGTATTTTCCCTTATTTTAG

#### 25 Intron 10 (SEQ ID NO 14)

GTGAGGCCCGTGCCGTGTCTCTGGGGGACCTCCACAGCCTGTGGGCTTTGCAGTTGAGCCCCCCGTGTCCTGCCCCTGG ACACACGTGGTGAGTGCAGGCGGTGACCTGGCTCCTGCTGCTCTTTGGAAAGTCAAGAGTGGCGGCTCCTGGGGCCCCAG TGAGACCCCCAGGAGCTGTGCACAGGGCCTGCAGGGCCGAGGCGCAGCCTCCCCCAGGGTGCACCTGAGCCTGCGGA GAGCAGGAGCTGCTGAGTGAGCTGGCCCACAGCGTTCGCTGCGGTCACGTTCCTGCGTGGGGTTGTTTGGGATCGGTGGG AGAATTTGGATTTGCTGAGTGCTGCTGTCTTGAACCACGGAGATGGCTAGGAGTGGGTTTCAGAGTTGATTTTTGTGAAT CAAACT'AAAATCAGGCACAGGGGACCTGGCCTCAGCACAGGGGATTGTCCAATGTGGTCCCCCTCAAGGGCGCCCCACAG AGCCGGTGGGCTTGTTTTAAAGTGCGATTTGACGAGGGACGAGAAACCTTGAAAGCTGTAAAGGGAACCCTCAGAAAATG TGGCCGCCAGGGGTGGTTTCAGGTGCTTTGCTGGGCTGTTTTGTGAAAACCCATTTGGACCCGCCCTCCAAGTCCACCC TGTGCACATTTAAATCCACTAAGATTCACTCGGGGGGAGCCCAGGTCCCAAGCAACTGAGGGCTCAGGAGTCCTGAGGCT GCTGAGGGGACAGACGGGGAACGCTGCTTCTGTGTGGCAAGTTCCTGAGGGTGCTGGCCAGGGAGGTGGTCAGA GTGTATGTTGGGGTCCCACCGGGGGCAGAACTCTGTCTCTGATGAGTCGGCAGCCATGTAACAGGAAGGGGTGGCCACAG GGAGCTGGGAATGCACCAGGGGAGCTGCGCAGCTGGCCGAGGTCCCAGGGCCAGGCCACAGGAAGGGCAGGGGACGCCC GGGGCCACAGCAGAGGCCGCAGGAAGGGAAGGGGATGCCCAGGCCAGAGCAGAGGCTACCGGGCACAGGGGGGCTCCCTG AGCTGGGTGAGCGAGGCTCATGACTCGGCGAGGGAACCTCCTTGACGTGAAGCTGACGACTGGTGTTGCCCAGCTCACAG CCCAGCCAGGTCCCGCGCCTGAGCAGGAACTCAGAACCCTCCCCTTTGTCTAAAGCACAGCAGATGCCTTCAGGGCATCT AGGAGAAAACAGGCAAAGTCGTTGAGAAACGTCTTAAAAGAAGGTGGGGATGGTGGCAATTTCTTGTCCAGATTTTAGTCT GCCCGGACCACAGATGAGTCTATAACGGGATTGTGGTGTTGCCATGGGGACACATGAGATGGACCATCACAGAGGCCAC 

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GGGAGACAGGGAAAGCACCCCGAAGTCTGGAGCAGGGCTGGGTCCAGGCTCCTCAGAGCTCCTGCCAGGCCCAGCACCCT
GCTCCAAATCACCACTTCTCTGGGGTTTTCCAAAGCATTTAACAAGGGTGTCAGGTTACCTCCTGGGTGACGGCCCCGCA
TCCTGGGGCTGACATTGCCCCTCTGCCTTAG

## 5 Intron 11 (SEQ ID NO 15)

GTGAGCGCACCTGGCCGGAAGTGGAGCCTGTGCCCGGCTGGGGCAGGTGCTGCAGGGCCGTTGCGTCCACCTCTGCT TCCGTGTGGGGCAGGCGACTGCCAATCCCAAAGGGTCAGAGGCCACAGGGTGCCCCTCGTCCCATCTGGGGCTGAGCAGA AATGCATCTTTCTGTGGGAGTGAGGGTGCTCACAACGGGAGCAGTTTTCTGTGCTATTTTGGTAAAAGGAAATGGTGCAC CAGACCTGGGTGCACTGAGGTGTCTTCAGAAAGCAGTCTGGATCCGAACCCAAGACGCCCGGGCCCTGCTGGGGCGTGAGT CTCTCAAACCCGAACACGGGGCCCTGCTGGGCATGAGTCCCTCTGAACCCGAGACCCTGGGGCCCTGCTGGGCGTGAGT CTCTCCGAACCCAGAGACTTCAGGGCCCTTTTGGGCGTGAGTCTCTCCGCTGTGAGCCCCACACTCCAAGGCTCATCCAC AATTCTGGGGTCTTGTTTCCCCAGAGCCCGAGAGCTCAAGGCCCCGTCTCAGGCTCAGACACAATGAATTGAAGATGGA ATAATCCCAGCACTTTGGGAGGCCGAGGTGGGTGGATCACTTGAGGCCAGGAGTTTGAGGCCAACCTAACCAACATAGTG AAATTCCATTTCTACTTAAAAAATACAAAAATTAGCCTGGCCTGGTGGCACACGCCTGTAGTCCCCGCTATGCGGGAGGC TGAGGCAGGAGAATCATTTGAACCCAGGAGGCAGAGGTTGCAGTGAGCCGAGATCACACCACTGCACTCCAGCCTGGGCA ACAGAGTGAGACTTCATCTTAAAAAAAAAAAAAAAAAGTATCAGCATTCCAAAACCATAGTGGACAGGTGTTTTTTTATTC TGTCCTTCGATAATATTTACTGGTGCTGTGCTAGAGGCCGGAACTGGGGGTGCCTTCCTCTGAAAGGCACACCTTCATGG GAAGAGAAATAAGTGGTGAATGGTTGTTAAACCAGAGGTTTAAACTGGGGTCCTGTCGTTCTGAGTTAACAGTCCAGATC TGGACTTTGCCTCTTTCCAGAATGCTCCCTGGGGTTTGCTTCATGGGGGAGCAGCAGGTGTGGACACCCTCGTGATGGGG GAGCAGCAGGTGCAGACGCCCTCATGATGGGGGGAGTGGCAGGTGCAGACACCCTTGTGCATGGTGCCCAGCATGTCCCTG TTGCAGCTCCCCCACAAGGATGCCGGTCTCCTGTGCTCCCCACAGTCCCTGCTTCCCTCTCACAGCCTTACCTGGTC CTGGCCTCCACTGGCTTTGTCTGCATGATTTCCACATTTCCTGGGCTCCCAGCACCTCTTCGCCTCTCCCAGGCACCTCT GCAGTCCTGGCCATACCAGTCAGCTGTGAACTGTCCACTGCTTATTTTGCTCCCCATGAAATGTATTTTTTAGGACAGGC CCAGAATATTCTGTGCTCCCAAAGGCCACTTGGTCAGAGTGTGTGCTTGCAGAGGTGGCTCTAAAAGCTCAGCAGTGGAG GCAGTGGTTCGCCATACTCAGGGTGAACTCACATCCTCTGTGTCTGAAGTATACAGCAGAGGCTTGAAGGGCATCTGGGA GAAGAAACAGGCAAAATGATTAAGAAAAGTGAAAAAGGAAAAGTGGTAAGATGGGAATTTTCTTGTCCAGATTTTAGTC TCCCAAACCACAGCTCAGATGGTAGAATGTGGTCAGAACTGATGGACAGAACAATAGAACAAAACGGAAGCCCTATCTCT GACTGGAAGCAAATAAGTTGTGTCTTTACAGCATATACCAGAGCAGATTCTAGGTAGAAGAGGAGACACATGCAAACAAC ACCAGCAACAGAAATAAAACAAAAGACTCAAAGGGAAGGGAGGTGAACGTTCCCTGGTTTGGTGTTGGGGAAGGACACAC AGGGAGGCGGATGAAACCAGTGAGGCAACGGGCATTGCTTTCACTGCAGAGAAACTCAGCTTGCCTGAGCCACAGTGAAA GTTCTCCTAACCACCTGAGAGGTAGAGGAGGAAAGGCTCCAGGGGGAGCAGCCGCCCTTGGTCACCCAGCTGGCAAAGGGC ATGCATGATTGCAGCCTGGCCTCCTGCTCCGGGGCCCTTGCTCTGCCCGAGGACCCCACACAAGTCAGACCCATAGGCTC CTACCAGCAGCGTCAAAGAAATGCATGTGAAACTGACAGCGAGACCCATCCCTCAAAGAAACGCACGTGAAACTGATGGC GAGACCTGTCCCCATCCTCATGCTGGCTCCTTTTCTGGGCTTGCCAAGAGCCAGCATCAGGTTGAGGCAAGCTGGAAAG ACTTTTCTGGAAAGCAGCTTGTTTGCATGGAAGTCCTCACAATGTCCTGTGTCTTCCCAGTAATTCCACTTCTGAAGTGA CAAATACAGGGCTAAGGAGATATTATGCATCACAAAACTTGCTCTGCCATTAAACATTTTTCAAAGAATTTTTGAAGAAT GTTTAATGGCACAAAACGTTTATTTCAATGTAGCAGTGTTCAAAGCTGGATGTAAAAGAACACCCCCAGGAGCCTGCCG 

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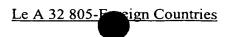
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#### Intron 12 (SEQ ID NO 16)

#### Intron 13 (SEO ID NO 17)

GTGAGCCGCCACCAAGGGGTGCAGGCCCAGCCTCCAGGGACCCTCCGCGCTCTGCTCACCTCTGACCCGGGGCTTCACCT TGGAACTCCTGGGTTTTAGGGGCAAGGAATGTCTTACGTTTTCAGTGGTGCTGCTGCCTGTGCACAGTTCTGTTCGCGTG GCTCTGTGCAAAGCACCTGTTCTCCATCTCTGGGTAGTGGTAGGAGCCGGTGTGGCCCCAGGTGTCCCCACTGTGCCTGT GCACTGGCCGTGGGACGTCATGGAGGCCATCCCAGGGCAGCAGGGGCATGGGGTAAAGAGATGTTTATGGGGAGTCTTAG CAGAGGAGGCTGGGAAGGTGTCTGAACAGTAGATGGGAGATCAGATGCCCGGAGGATTTGGGGTCTCAGCAAAGAGGGCC ACAACTTTATCACAGAGGGAAGGGCCAATCTGTGGAGGCCACAGGGCCAGCTTCTGCCTGGAGTCAGGGCAGGTGGTGGC ACAAGCCTCGGGGCTGTACCAAAGGGCAGTCGGGCACCACAGGCCCGGGCCTCCACCTCAACAGGCCTCCCGAGCCACTG GGAGCTGAATGCCAGGAGGCCGAAGCCCTCGCCCCATGAGGGCTGAGAAGGAGTGTGAGCATTTGTGTTACCCAGGGCCG AGGCTGCGCGAATTACCGTGCACACTTGATGTGAAATGAGGTCGTCGTCTATCGTGGAAACCCAGCAAGGGCTCACGGGA GAGTTTTCCATTACAAGGTCGTACCATGAAAATGGTTTTTAACCCGAGTGCTTGCGCCTTCATGCTCTGGCAGGAGGGC AGAGCCACAGCTGCATGTTACCGCCTTTGCACCAGCTCCAGAGGCTTGGGACCAGGCTGTCTCAGTTCCAGGGTGCGTCC GGCTCAGACCGCCCTCCTCTCTCCTCTCTCTCTCCTCAAATCTTCCCTCGTTTTGCATCTCCCTGACGCGTGCCTGGG CCCTCGTGCAAGCTGCTTGACTCCTTTCCGGAAACCCTTGGGGTGTGCTGGATACAGGTGCCACTGAGGACTGGAGGTGT CAGGTGAAAACTCCTGGGAAACTCCCAGGGCCATGTGACCTGCCACCTGCTCCCATATTCAGCTCAGTCTTGTCCTC ATTTCCCCACCAGGGTCTCTAGCTCCGAGGAGCTCCCGTAGAGGGCCTGGGCTCAGGGCAGGGCGGCTGAGTTTCCCCAC CCATGTGGGGACCCTTGGGTAGTCGCTTGATTGGGTAGCCCTGAGGAGGCCGAGATGCGATGGGCCACGGGCCGTTTCCA AACACAGAGT JAGGCACGTGGAAGGCCCAGGAATCCCCTTCCCTCGAGGCAGGAGTGGGAGAACGGAGAGCTGGGCCCCG ATTTCACGGCAGCCAGGCTGCAGTGGCCGAGGCTGTGGTCCACGTGGCGCTGGGGGCGGGGTCTGATTCAAATCCGC TGGGGCTCGGCCTGCCGGCCGCGCGCCCCCCACACGGGCTTGGGGTGGACGCCCCGACCTCTAGCAGGTGGC TATTTCTCCCTTTGGAAGAGCCCCTCACCCATGCTAGGTGTTTCCCTCCTGGGTCAGGAGCGTGGCCGTGTGGCAACC

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CCGGGACCTTAGGCTTATTTATTTGTTTAAAAACATTCTGGGCCTGGCTTCCGTTGTTGCTAAATGGGGAAAAGACATCCCACCTCAGCAGAGTTACTGAGAGGCTGAAACCGGGGTGCTGGCTTGACTGGTGTGATCTCAGGTCATTCCAGAAGTGGCT CAGGAAGTCAGTGAGACCAGGTACATGGGGGGGCTCAGGCAGTGGGTGAGATGAGGTACACGGGGGGCTCAGGCAGTGGGT GAGGCCAGGTACATGGGGGGGCTCAGGCACTGGGTGAGATGAGGTACACGGGGGGGCTCAGGCAGAGGGTCAGACCAGGTAC ACGGGGGCTCTGATCACACGCACATATGAGCACATGTGCACATGTGCTGTTTCATGGTAGCCAGGTCTGTGCACACCTGC CCCAAAGTCCCAGGAAGCTGAGAGGCCAAAGATGGAGGCTGACAGGGCTGGCGCGGTGGCTCACACCTGTAGTCCCAGCA  $\tt CTTTGGGAGGCCGAGGGGATCCCTTGAGCCCAGGAGTTTAAGACCAGCCTGAGCAACATAGTAGAACCCCATCTC$ TATGAAAAATAAAAACTAACTGAACATGGTGGTGTGCGCCTGTAGTTCCAATACTTGGGAGGCTGAAGTGGGAG GATCACTTGAGCCCAGGAGGTGGAAGCTGCAGTGAGCTGAGATTGCACCACTGTACTGCAGCCTGGGTGACAGAGTGAGA GCCCATCTCAACAACAACAAGAAGACTGACAAATGCAGTTTCTTGGAAAGAACATTTAGTAGGAACTTAACCTACACA CACCACAGGGGGGGGGGGCTCAGAAGGGATGCGCAGGACGTTGATATACGATGACATCAAGGTTGTCTGACGAAGGGCAG GATTCATGATAAGTACCTGCTGCTACACAAGGAACAATGGATAAACTGGAAACCTTAGAGGCCTTCCCGGAACAGGGGCCT  ${\tt AATCAGAAGCCAGCATGGGGGGCTGGCATCCAGGATGGAGCTGCTTCAGCCTCCACATGCGTGTTCATACAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGAGCAGCATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCACATGCAGATGGTGCAGATGGTGCAGATGGTGCAGATGCAGATGGTGCAGATGCAGATGGTGCAGATGCAGATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATG$ GCCCACACCCACGAGCACCGTCTGATTAGGAGGCCTTTCCTCTGACGCTGTCCGCCATCCTCTCAG

#### Intron 14 (WEQ ID NO 18)

#### Intron 15 (SEQ ID NO 19)

#### 40 3'-untranscribed region (SEQ ID NO 20)

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CGGGGAAGATGGGGAAGCCTGGCTGGGCCCCTCCTCCCCTGCCTCCACCTGCAGCCGTGGATCCGGATGTGCTTCCCT GGTGCACATCCTCTGGGCCATCAGCTTTCATGGAGGTGGGGGGCAGGGGCATGACACCATCCTGTATAAAATCCAGGATT CCTCCTCCTGAACGCCCCAACTCAGGTTGAAAGTCACATTCCGCCTCTGGCCATTCTCTTAAGAGTAGACCAGGATTCTG ATCTCTGAAGGGTGGGTAGGGTGGGGCAGTGGAGGGTGTGGACACAGGAGGCTTCAGGGTGGGGCTGGTGATGCTCTCTC ATCCTCTTATCATCTCCCAGTCTCATCTCTCATCCTCTTATCATCTCCCAGTCTCATCTGTCTTCCTCTTATCTCCCAGT CTCATCTGTCATCCTCTTACCATCTCCCAGTCTCATCTCTTATCCTCTTATCTCCTAGTCTCATCCAGACTTACCTCCCA GAGGGGCGGCTCAGAGGGACGCAGTCTTGGGGTGAAGAAACAGCCCCTCCTCAGAAGTTGGCTTGGGCCACACGAAACCG AGGGCCCTGCGTGAGTGGCTCCAGAGCCTTCCAGCAGGTCCCTGGTGGGGCCTTATGGTATGGCCGGGTCCTACTGAGTG CACCTTGGACAGGGCTTCTGGTTTGAGTGCAGCCCGGACGTGCCTGGTGTCGGGGTGGGGGCTTATGGCCACTGGATATG GCGTCATTTATTGCTGCTGCTTCAGAGAATGTCTGAGTGACCGAGCCTAATGTGTATGGTGGGCCCAAGTCCACAGACTG GCGCCTTTGCCCTGCAAACTGGAAGGGAGCGGCCCCGGGCGCCCTTGGGCGACGACCTCAAGTGAGAGGTTGGACAGAAC AGGGCGGGGACTTCCCAGGAGCAGAGGCCGCTGCTCAGGCACACCTGGGTTTGAATCACAGACCAACaGGTCAGGCCATT GTTCAGCTATCCATCTTCTACAAAGCTCCAGATTCCTGTTTCTCCGGGTGTTTTTTGTTGAAATTTTACTCAGGATTACT TATATTTTTTGCTAAAGTATTAGACCCTTAAAAAAGGTATTTGCTTTGATATGGCTTAACTCACTAAGCACCTACTTTAT TTGTCTGTTTTTATTATTATTATTATTATTATTATTAGAGATGGTGTCTACTCTGTCACCCAGGTTGTTAGTGCAGTGGCAC AGTCATGGCTCGCTGTAGCCGCAAACCCCCAGGCTCAAGTGATCCTCCGGCCTCAGCTTCCCAGAGTGCTGGGATTACAG GTGTGAGCCACTGCCCTTGCCTGGCACTTTTAAAAACCACTATGTAAGGTCAGGTCCAGTGGCTTCCACACCTGTCATCC CAGTAGTTTGGGAAGCCGAGGCAGAAGGATTGTCTGAGGCCAGGAGTTTGAGACCAGCATGGGTAACATAGGGAGACCCC ATCTCTACAAAAATGCAAAAAGTTATCCGGGCGTGGGGTCCAGCATCTGTAGTCCCAGCTGCTCGGGAGGCTGAGTGGG AGGATCGCTTGAGCCCGGGAGGTCATGGCTGCAGTGAGCTGTGATTGTACCATCGCACTCCAGCCTGGGCAACAGAGTGA GAAGAAGGAAGAAGAAGGAGGAGGAGGCCTGCTAGGTGCTAGGTAGACTGTCAAATCTCAGAGCAAAATGAAAATAACA GACAAGCGTGTATGGAGCGAGTGAGTTCAAAGCAGAAAGGGAGGAGAAGCAGGCAAGGGTGGAGGCTGTGGGTGACACCA GCCAGGACCCCTGAAAGGGAGTGGTTGTTTTCCTGCCTCAGCCCCACGCTCCTGCCGGTCCTGCACCTGCTGTAACCGTC GATGTTGGTGCCAGGTGCCCACCTGGGAAGGATGCTGTGCAGGGGGGCTTGCCAAACTTTGGTGGGTTTCAGAAGCCCCAG GCACTTGTGGCAGGCACAATTACAGCCCCTCCCCAAAGATGCCCACGTCCTTCTCCTGGAACCTGTGAATGTGTCACCCG CAAGGCAGAGGCTGGTGAAGGCTGCAGGTGGAATCACGGCTGCCAGTCAGCCGATCTTAAGGTCATCCTGGATTATCTGG TGGGCCTGATATGGCCACAAGGGTCCCTAGAAGTGAGAGAGGGGAGGCAGGGGAGAGTCAGAGAGGGGACGTGAGAAGGAC CACTGGCCACTGCTGGCTTTGAGATGGAGGAGGGGGTCCCCAGCCAAGGAATGGGGGCAGCCGCTCCATGCTGGAAAAGC AAGCAATCCTCCCGGTCCTGAGGGCACACGGCCCTGCCCACGCCTCGATTTCAGGCCAGTGGGACCTGTTTCAGCTTTC CGGCCTCCAGAGCTGTAAGATGATGCGTTTGTGTTCAGCCACTAAGCTGCAGTGATTCGTCACAGCAGCAAATGGAATAG

CAGTACAGGGAAATGAATACAGGGACAGTTCTCAGAGTGACTCTCAGCCCACCCCTGGG

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Characterization of the exons showed, interestingly, that the functionally important hTC protein domains which are described in our Patent Application PCT/EP/98/03469 are arranged on separate exons. The telomerase-characteristic T motif is located on exon 3. The RT (reverse transcriptase) motifs 1-7, which are important for the catalytic function of the telomerase, are located on the following exons: RT motifs 1 and 2 on exon 4, RT motif 4 on exon 9, RT motif 5 on exon 10, and RT motifs 6 and 7 on exon 11. RT motif 3 is shared by exons 5 and 6 (see Fig. 8).

Elucidation of the exon-intron structure of the hTC gene also shows that the four deletions or insertion variants of the hTC cDNA which were described in our Patent Application PCT/EP/98/03469, as well as three additional hTC insertion variants which are described in the literature (Kilian et al., 1997), in all probability represent alternative splicing products. As shown in Fig. 8, the splicing variants can be divided into two groups: deletion variants and insertion variants.

The hTC variants in the deletion group lack specific sequence segments. The 36 bp in-frame deletion in variant DEL1 in all probability results from using an alternative 3' splice acceptor sequence in exon 6, resulting in a part of RT motif 3 being lost. In variant DEL2, the normal 5' splice donor and 3' splice acceptor sequences of introns 6, 7 and 8 are not used. Instead exon 6 is fused directly to exon 9, resulting in a displacement arising in the open reading frame and a stop codon appearing in exon 10. Variant Del3 is a combination of variants 1 and 2.

The insertion variant group is characterized by the insertion of intron sequences which lead to premature cessation of translation. Instead of the 5' splice donor sequence of intron 5, which is normally used, use is made, in variant INS1, of an alternative, 3'-located splice site, resulting in the insertion of the first 38 bp from intron 4 between exon 4 and exon 5. The insertion, in variant INS2, of a region of the intron 11 sequence likewise results from using an alternative 5' splice donor sequence in intron 11. Since this variant was only described inadequately in the

literature (Kilian et al., 1997), it is not possible to determine the precise alternative 5' splice donor sequence in this variant. The insertion of intron 14 sequences between exon 14 and exon 15 in variant INS3 comes from using an alternative 3' splice acceptor sequence, resulting in the 3' part of intron 14 not being spliced.

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The hTC variant INS4 (variante 4), which is described in our Patent Application PCT/EP/98/03469, is characterized by exon 15, and the 5' part region of exon 16, being replaced by the first 600 bp of intron 14. This variant can be attributed to the use of an alternative internal 5' splice donor sequence in intron 14 and an alternative 3' splice acceptor sequence in exon 16, resulting in an altered C terminus.

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The *in vivo* generation of hTC protein variants which are probably non-functional and which could interfere with the function of the complete hTC protein constitutes a possible mechanism, in addition to transcription regulation, for controlling hTC protein function. The function of the hTC splicing variants is not yet known. Although most of these variants presumably encode proteins without reverse transcriptase activity, they could nevertheless play a crucial role as transdominant-negative telomerase regulators by, for example, competing for interaction with important binding partners.

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The search for possible transcription factor binding sites was carried out using the "find pattern" algorithm from the Genetics Computer Group (Madison, USA) GCG Sequence Analysis program package. This resulted in the identification of a variety of potential binding sites for transcription factors in the nucleotide sequence of intron 2, which binding sites are listed in Tab. 2. In addition, an Sp1 binding site was found in intron 1 (pos. 43), and a c-Myc binding site was found in the 5'-untranslated region (cDNA position 29-34, cf. Fig. 6).

## Example 6

In order to ascertain the start point(s) of hTC transcription in HL 60 cells, the 5' end of the hTC mRNA was determined by means of primer extension analysis.

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2 μg of polyA<sup>+</sup> RNA from HL-60 cells were denaturated at 65°C for 10 min. 1 μl of RNasin (30-40 U/ml) and 0.3-1 pmol of radioactively labelled primer (5'GTTAAGTTGTAGCTTACACTGGTTCTC 3'; 2.5-8x10<sup>5</sup> cpm) were added for primer annealing, and the whole was incubated, at 37°C for 30 min, in a total volume of 20 µl. After the addition of 10 µl of 5xreverse transcriptase buffer (from Gibco-BRL), 2 µl of 10 mM dNTPs, 2 µl RNasin (see above), 5 µl of 0.1 M DTT (from Gibco-BRL) 2 µl of ThermoScript RT (15 U/µl; from Gibco-BRL) and 9 µl of DEPC-treated water, primer extension took place, at 58°C for 1 h, in a total volume Ilacuna]. The reaction was stopped by adding 4 µl of 0.5 M EDTA, pH 8.0, and the RNA was degraded, at 37°C for 30 min, after having added 1 µl of RNaseA (10 mg/ml), 2.5 µg of sheared calf thymus DNA and 100 µl of TE were then added, and the mixture was extracted once with 150 µl of phenol/chloroform (1:1). The DNA was precipitated, at -70°C for 45 min, after adding 15 µl of 3 M Na acetate and 450 µl of ethanol, and then centrifuged at 14,000 rpm for 15 min. The precipitate was washed once with 70% ethanol, dried in air and dissolved in 8 µl of sequencing stop solution. After 5 min of denaturation at 80°C, the samples were loaded onto a 6% polyacrylamide gel and fractionated electrophoretically (Ausubel et al., 1987) (Fig. 5).

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In this connection, a main transcription start site was identified which is located 1767 bp 5' of the ATG start codon of the hTC cDNA sequence (nucleotide position 3346 in Fig. 4). In addition to this, the nucleotide sequence around this main transcription start (TTA<sub>+1</sub>TTGT) represents an initiator element (Inr), which, in 6 out of 7 nucleotides, matches the consensus motif (PyPyA<sub>+1</sub>Na/tPyPy) (Smale, 1997) of an initiator element.

It was not possible to identify any unambiguous TATA box in the immediate vicinity of the experimentally identified main transcription start, which means that the hTC promoter has probably to be classified in the family of TATA-less promoters (Smale, 1997). However, a potential TATA box from nucleotide position 1306 to nucleotide position 1311 (Fig. 4) was found by means of bioinformatics analysis. The subsidiary transcription starts which were additionally observed around the main transcription start have also been described in the case of other TATA-less promoters (Geng and Johnson, 1993), for example in the strongly regulated promoters of some cell cycle genes (Wick *et al.*, 1995).

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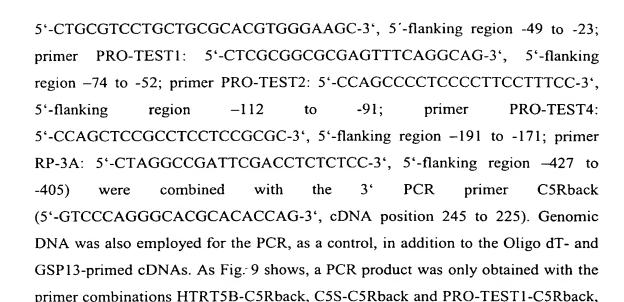
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## Example 7

In addition to the start point of the hTC transcript which was described in Example 6 and identified in HL60 cells, a further transcription start region was also identified in HL60 cells. With the aid of RT-PCR analyses, the region of the hTC gene transcription start in HL60 cells was localized to bp -60 to bp -105.

The cDNA for this was synthesized using a First Strand cDNA Synthesis kit (Clontech), in accordance with the manufacturer's instructions, and employing 0.4 µg of HL60 cell polyA RNA (Clontech) and the gene-specific primer GSP13 (5'-CCTCCAAAGAGGTGGCTTCTTCGGC-3', cDNA position 920-897). In a final volume of 50 µl, 10 pmol dNTP mix were added to 1 µl of cDNA, and a PCR reaction was carried out in 1xPCR reaction buffer F (PCR-Optimizer kit from InVitrogen) and using one unit of platinum Taq DNA polymerase (from Gibco/BRL). 10 pmol of each of the 5' and 3' primers defined below were added as primers. The PCR was carried out in 3 steps. A two-minute denaturation at 94°C was followed by 36 PCR cycles in which the DNA was first of all denatured at 94°C for 45 sec and, after that, the primers were annealed, and the DNA chain was extended at 68°C for 5 min. The cycles were concluded by a chain extension at 68°C for 10 min. In all, six 5' HTRT5B: **PCR** (primer different primers 5'-CGCAGCCACTACCGCGAGGTGC-3', cDNA position 105 to 126; primer C5S: ign Countries

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indicating that the start point for hTC transcription lies in the region between bp-60

## Example 8

and bp-105.

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Several extremely GC-rich regions, so-called CpG Islands, are located in the isolated 5'-flanking region, of about 11.2 kb in size, of the hTC gene. One CpG Island, having a GC content of > 70%, extends from bp - 1214 into intron 2. Two further GC-rich regions having a GC content of > 60% extend from bp - 3872 to bp - 3113 and from bp - 5363 to bp - 3941, respectively. The positions of the CpG Islands are shown graphically in Fig. 11.

The search for possible transcription factor binding sites was carried out using the "Find Pattern" algorithm from the Genetics Computer Group (Madison, USA) GCG Sequence Analysis program package. This resulted in the identification of a variety of potential binding sites in the region up to -900 bp upstream of the translation start codon ATG: five Sp1 binding sites, one c-Myc binding site, and one CCAC box (Fig. 10). In addition, a CCAAT box and a second c-Myc binding site were found at positions –1788 and –3995, respectively, of the 5'-flanking region.



## Example 9

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In order to analyse the activity of the hTC promoter, PCR amplification was used to generate four hTC promoter sequence segments of differing length, which segments were cloned into the Promega vector pGL2 5' in front of the luciferase reporter gene. The 8.5 kb SacI fragment which was subcloned from phage clone P12 was selected as the DNA source for the PCR amplification. In a final volume of 50 µl, 10 pmol of dNTP mix were added to 35 ng of this DNA, and a PCR reaction was carried out in 1xPCR reaction buffer (PCR-Optimizer kit from InVitrogen) and using one unit of platinum Taq DNA polymerase (from Gibco/BRL). In each case 20 pmol of the 5' and 3' primers which are defined below were added as primers. The PCR was carried out in three steps. A two-minute denaturation at 94°C was followeed by 30 PCR cycles in which the DNA was first of all denaturated at 94°C for 45 sec, after which the primers were annealed, and the DNA chain was extended, at 68°C for 5 min. The cycles were concluded by a chain extension at 68°C for 10 min. The selected 3' PCR the primer PK-3A primer each case was (5'-GCAAGCTTGACGCAGCGCTGCCTGAAACTCG-3', position -43 to -65), which primer recognizes a sequence region 42 bp upstream of the ATG START codon. A promoter fragment of 4051 bp in size (NPK8) was amplified by combining the PK-3A primers with the 5' PCR primer PK-5B (5'-CCAGATCTCTGGAACACAGAGTGGCAGTTTCC-3', position -4093 to PK-5C -4070). Combining the pair of primers PK-3A and (5'-CCAGATCTGCATGAAGTGTGTGGGGATTTGCAG-3', position -3120 to -3096) led to the amplification of a promoter fragment of 3078 bp in size (NPK15). of and PK-5D Use the primer combination PK-3A (5'-GGAGATCTGATCTTGGCTTACTGCAGCCTCTG-3', position –2110 -2087) amplified a promoter fragment of 2068 bp in size (NPK22). Finally, using the PK-5E PK-3A and primer combination (5'-GGAGATCTGTCTGGATTCCTGGGAAGTCCTCA-3', position -1125 to -1102) led to the amplification of a promoter fragment of 1083 bp in size (NPK27).

The PK-3A primer contains a HindIII recognition sequence. The different 5' primers contain a BglII recognition sequence.

The resulting PCR products were purified using the Qiagen QIA quick spin PCR purification kit, in accordance with the manufacturer's instructions, and then digested with the restriction enzymes BgIII and HindIII. The pGL2 promoter vector was digested with the same restriction enzymes, and the SV40 promoter contained in this vector was released and removed. The PCR promoter fragments ligated into the vector, which was then transformed into competent DH5α bacteria (from Gibco/BRL). DNA for the promoter activity analyses, which are described below, was isolated from transformed bacterial clones using the Qiagen plasmid kit.

#### Example 10

The activity of the hTC promoter was analysed in transient transfections in eukaryotic cells.

All the work with eukaryotic cells was carried out at a sterile workstation. CHO-K1 and HEK 293 cells were obtained from the American Type Culture collection.

CHO-K1 cells were kept in DMEM Nut Mix F-12 cell culture medium (from Gibco-BRL, order number: 21331-020) containing 0.15% streptomycin/penicillin, 2 mM

glutamine and 10% FCS (from Gibco-BRL).

HEK 293 cells were cultured in DMOD cell culture medium (from Gibco-BRL, order number: 41965-039) containing 0.15% streptomycin/penicillin, 2 mM glutamine and 10% FCS (from Gibco-BRL).

CHO-K1 and HEK 293 cells were cultured at 37°C in a water-saturated atmosphere while being gassed with 5% CO<sub>2</sub>. When the cell lawn was confluent, the medium was sucked off, after which the cells were washed with PBS (100 mM KH<sub>2</sub>PO<sub>4</sub> pH

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7.2; 150 mM NaCl) and released by adding a trypsin-EDTA solution (from Gibco-BRL). The trypsin was inactivated by adding medium and the cell count was determined using a Neubauer counting chamber in order to plate out the cells at the desired density.

For the transfection, in each case 2x 10<sup>5</sup> HEK 293 cells were plated out, per well, in a 24-well cell culture plate. The HEK 293 medium was removed after 3 hours. For the

transfection, up to 2.5  $\mu g$  of plasmid DNA, 1  $\mu g$  of a CMV  $\beta\text{-}Gal$  plasmid construct

(from Stratagene, order numner: 200388), 200  $\mu l$  of serum-free medium and 10  $\mu l$  of

transfection reagent (DOTAP from Boehringer Mannheim) were incubated at room

temperature for 15 minutes and then dropped uniformly onto the HEK 293 cells. 1.5

ml of medium were added after 3 hours. The medium was changed after 20 hours.

After a further 24 hours, the cells were harvested for determining the luciferase activity and the β-Gal activity. For this, the cells were lysed, at room temperature for

15 minutes, in the cell culture lysis reagent (25 mM Tris [pH 7.8] containing H<sub>3</sub>PO<sub>4</sub>;

2 mM CDTA; 2 mM DTT; 10% glycerol; 1% Triton X-100). Twenty μl of this cell

lysate were mixed with 100 µl of luciferase assay buffer (20 mM Tricin; 1.07 mM

(MgCO<sub>3</sub>)<sub>4</sub> Mg(OH)<sub>2</sub>·5H<sub>2</sub>O; 2.67 mM MgSO<sub>4</sub>; 0.1 mM EDTA; 33.3 mM DTT;

270 μM coenzyme A; 470 μM luciferin, 530 μM ATP), and the light generated by

20 the luciferase was measured.

420 nm.

In order to measure the  $\beta$ -galactosidase activity, equal quantities of cell lysate and  $\beta$ -galactosidase assay buffer (100 mM sodium phosphate buffer, pH 7.3; 1 mM MgCl<sub>2</sub>; 50 mM  $\beta$ -mercaptoethanol; 0.665 mg of ONPG/ml) were incubated at 37°C for at least 30 minutes or until a slight yellow coloration appeared. The reaction was stopped by adding 100  $\mu$ l of 1 M Na<sub>2</sub>CO<sub>3</sub>, and the absorption was determined at

In order to analyse the hTC promoter, four hTC promoter sequence segments of differing length were cloned 5' in front of the luciferase reporter gene (cf. Example 9).

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The relative luciferase activities of two independent transfections in HEK 293 cells, using the constructs NPK8, NPK15, NPK22 and NPK27, are plotted in Fig. 11. Each experiment was carried out in duplicate. The standard deviation has also been given. The construct NPK 27 exhibits a luciferase activity which is 40 times higher than the basal activity of the promoterless luciferase control construct (pGL2-basic) and from 2 to 3 times higher than that of the SV40 promoter control construct (pGL2PRO). Interestingly, a luciferase activity which was from 2 to 3 times lower than that obtained with the NPK 27 construct was observed in cells which were transfected with longer hTC promoter constructs (NPK8, NPK15, NPK22). Similar results were also observed in CHO cells (data not shown).



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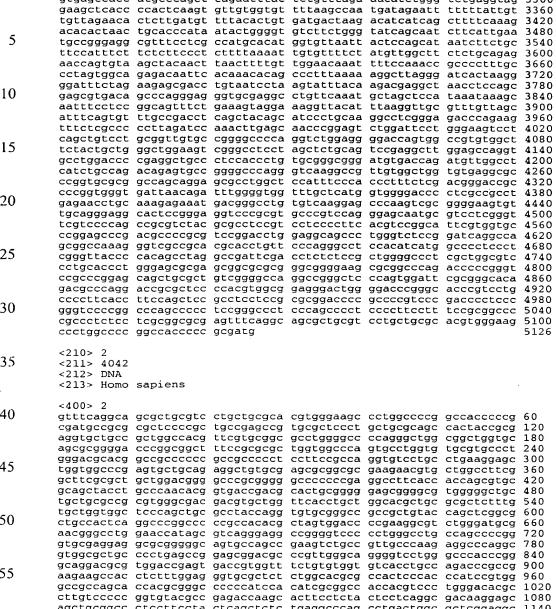
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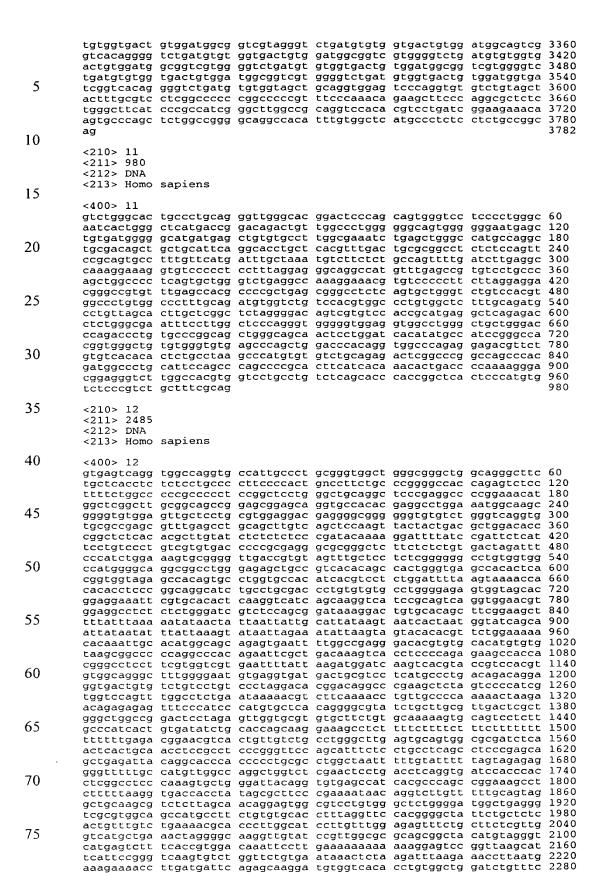
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- 74 -

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